

# **SPECTROPHOTOMETRY**

# SPECTROPHOTOMETRY

- It is that technique that measures the amount of light absorbed or transmitted by a substance.
- It is an analytical technique that is applied to obtain valuable information, such as:
  - The identity of unknown compounds by their characteristic absorption spectra in the ultraviolet, visible or infrared regions (qualitative analysis) .
  - Determination of the unknown concentration of an analyte by measuring the light absorption at one or more wavelengths (quantitative analysis) .
- Enzyme catalyzed reactions can be followed by measuring the absorption of the substrate or product .

# REGIONS OF THE ELECTROMAGNETIC SPECTRUM

Regions	X-Ray	Ultraviolet	Visible	Infrared	microwave
Wave length	0.1 -100 nm	100 - 400 nm	400 - 800 nm	800 nm - 100 $\mu\text{m}$	100 $\mu\text{m}$ - 30 cm

The ultra violet and the visible regions are the ones that we usually use in the spectrophotometry.

$$1 \text{ nm} = 10^{-9} \text{ m}$$

$$1 \mu\text{m} = 10^{-6} \text{ m}$$

$\lambda$  is the symbol of wavelength.

# THE ESSENTIAL COMPONENTS OF SPECTROPHOTOMETER

## 1- Light source:

- It can be two kinds:
  - Tungsten lamp ; produces light at visible region.
  - Hydrogen lamp; produces light at ultraviolet region.

## 2- Collimator:

- It is a focusing device that transmits an intense straight beam of light.

## 3- Monochromator:

- It is a device that divides the light beam into it's component wavelengths.

# THE ESSENTIAL COMPONENTS OF SPECTROPHOTOMETER

## 4- Selector:

- It selects the required wavelength.

## 5- Cuvette:

- It is a compartment in which the sample is placed.
- Three kinds:
  - Glass and plastic cuvettes; used in the visible region.
  - Quartz cuvettes; used in the ultraviolet region,

The glass cuvettes absorb light in the ultraviolet region → Thus the amount of light measured by spectrophotometer will be the absorbance of sample + the glass cuvette.

# THE ESSENTIAL COMPONENTS OF SPECTROPHOTOMETER

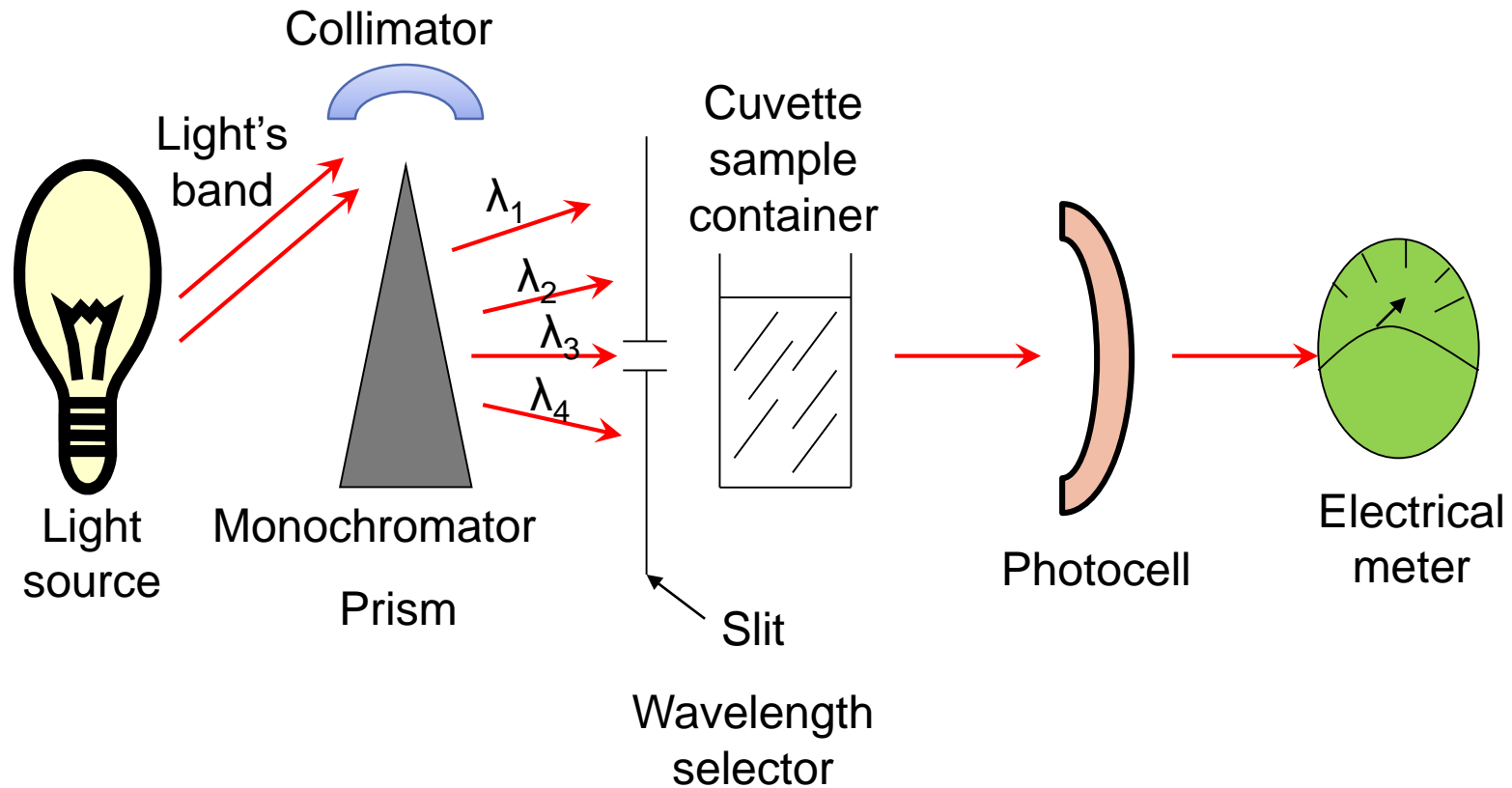
## 6- Photocell (photodetector):

- It detects the amount of light transmitted.

## 7- Electrical meter:

- It records the output of the detector.

# THE ESSENTIAL COMPONENTS OF SPECTROPHOTOMETER



# **BEER and LAMBERT LAWS**

**The fraction of the incident light  $I_0$  that is absorbed by a solution depends on three factors:**

- 1- The thickness of the sample or path length.
- 2- The concentration of the absorbing sample.
- 3- The chemical nature of the compound.

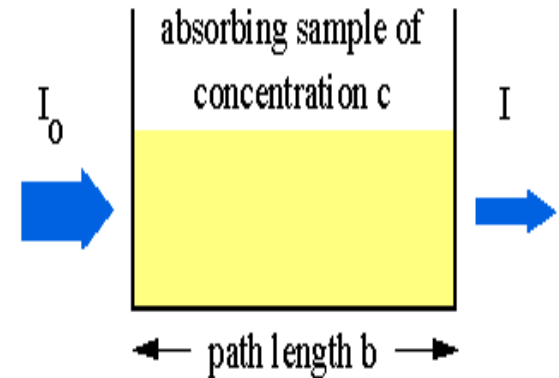
**The relationship between the concentration, path length, and the amount of light absorbed or transmitted can be exposed mathematically in two laws: the Beer law and Lambert law.**



# BEER'S LAW

- It states that the intensity of the light transmitted by an absorbing media decreases with increasing concentration of the absorbing compound.

$$\text{Log } I_0 / I \propto c$$



$I_0$  = intensity of the incident light

$I$  = intensity of the transmitted light

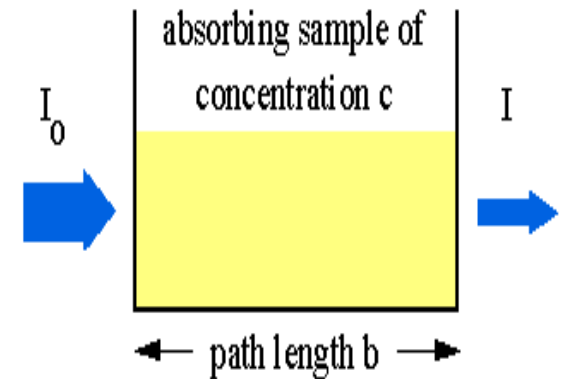
$c$  = concentration of absorbing compound

- The amount of light absorbed =  $I_0 - I$
- $\text{Log } I_0 / I$  represents the fraction of light absorbed

# LAMBERT'S LAW

It states that the intensity of the light transmitted by an absorbing media decreases as the thickness or path length of the absorbing media increases.

$$\text{Log } I_0 / I \propto L$$



$L$  = is the path length

# BEER-LAMBERT LAW

The two law's can be combined in one law which is Beer-Lambert Law

$$A = \text{Log } I_0 / I = a c L$$

**A** = absorbance or optical density O.D.

**a** = is the **extinction coefficient** which is a constant for each substance but varies at different wavelength

If the concentration is expressed in :

- in Molaity → Molar absorption coefficient ( $a_m$ )
- in g/L → Specific absorption coefficient ( $a_s$ )
- $a_{m340}$  = is the molar absorption coefficient of a substance at a wavelength = 340 nm

$$a_m = a_s Mwt$$

$a_m$  is most commonly used in biochemistry, and the path length  $L$  is almost always 1 cm, thus the units for  $a_m$  is  $M^{-1} \text{ cm}^{-1}$ .

# BLANK SOLUTION

Is a solution that is necessary in all spectrophotometry studies.

It should contain all components of the assay or test solution *except the component who's absorbance is being measured*.

## *Purpose of the Blank:*

The blank will cancel out the absorbance of the substances in the background so that the absorbance of the tests will be that of the compound under study only.

# **SOLUTIONS CONTAINING ONE ABSORBING SUBSTANCE**

## **Example 1:**

**A solution containing 2 g/l of a light absorbing substance in a 1 cm cuvette transmits 75% of the incident light at 260 nm. Calculate the transmission of a solution containing**

**a) 4 g/l.**

**b) 6 g/l.**

**c) If the Mwt is 250, calculate  $a_m$ .**

**d) What type of cuvette should you use here? Why?**

**First, Calculate the A , and a<sub>s</sub>**

$$A = \text{Log } I_0 / I$$

$$A = \log 1.0 / 0.75 = 0.124$$

$$A = a_s c l$$

$$a_s = A / c l = 0.124 / 2 \times 1$$

$$a_s = 0.0625 \text{ (g/L)}^{-1} \text{ cm}^{-1}$$

**a) transmission of a solution containing 4 g/l.**

$$\log I_0 / I = a_s c l$$

$$\text{Log } 1.0 - \log I = 0.0625 \times 4 \times 1$$

$$0 - \log I = 0.25$$

$$-\log I = 0.25 \text{ ( divide by -1) } \rightarrow \log I = -0.25$$

$$I = \text{antilog } (- 0.25 ) = 0.562 \text{ (multiply by 100) } \rightarrow 56.2\%$$

**Note:**

***I<sub>0</sub> is always 100%*** → as decimal (100/100= 1)

I = 75% → as decimal (75/100= 0.75)

**Absorbance (A) does not have a unit**

**b)**  $\text{Log } I_0 / I = a_s c l$

$\text{Log } 1.0 - \log I = 0.0625 \times 6 \times 1.$

$-\log I = 0.375$

$\text{Log } I = -0.375$

$I = \text{antilog } -0.375 = 0.422 \rightarrow 42.2\%$

**C)**  $a_m = a_s \times \text{Mwt} = 0.0625 \times 250 = 15.63 \text{ M}^{-1} \text{ cm}^{-1}$

**D)** Quartz cuvettes should be used at the U.V range.

## **EXAMPLE 2**

**A solution containing  $10^{-5}$  M ATP, has a transmission 0.702 (70.2%) at 260 nm in a 1 cm cuvette. Calculate:**

- a) The transmission of the solution in a 3 cm cuvette.**
- b) The absorbance of the solution in a 1 cm and 3 cm cuvette.**
- c) The absorbance if the concentration increased to  $5 \times 10^{-5}$  M of ATP, in a 1 cm cuvette.**



**First, Calculate the A , and  $a_m$**

$$A = \text{Log } I_0 / I = a_m c l$$

$$A = \log 1.0 / 0.702 = 0.152$$

$$0.152 = a_m \times 10^{-5} \times 1$$

$$a_m = 0.152 / 10^{-5} = 15200 \text{ M}^{-1} \text{ cm}^{-1}$$

**a) transmission of the solution in a 3 cm cuvette.**

$$A = 15200 \times 10^{-5} \times 3 = 0.456$$

$$A = \text{Log } I_0 / I$$

$$0.456 = \log 1.0 / I$$

$$0.456 = \log 1 - \log I = 0 - \log I = -\log I$$

$$\text{Thus } I = \text{antilog } -0.456 = 0.349 \rightarrow 34.9\%$$

**b) A in a 1 cm cuvette.**

$$A = 15200 \times 10^{-5} \times 1 = 0.15$$

**A in a 3 cm cuvette.**

$$A = 15200 \times 10^{-5} \times 3 = 0.456$$

**c) A if the concentration increased to  $5 \times 10^{-5} \text{ M}$  of ATP, in a 1 cm cuvette.**

$$A = 15200 \times (5 \times 10^{-5}) \times 1 = 0.76$$

# PROTEIN DETERMINATIONS

Proteins in solutions can be determined spectrophotometrically by two methods:

## a) Colorimetric method: (e.g. *Biuret method*)

it is based on the reaction of  $\text{Cu}^{2+}$  with peptides in an alkaline solution producing a purple complex that has an absorption maximum at 540 nm.



## b) Direct spectrophotometry:

The absorbance at **280nm** can be used to determine protein concentration in solutions.

(Because proteins have a distinct absorbance maximum at 280nm due to their **aromatic amino acids**).

## **EXAMPLE 3**

**A protein solution (0.3 ml) was diluted with 0.9 ml of water. To 0.5 ml of this diluted solution, 4.5 ml of biuret reagent was added and the color was allowed to develop.**

**The absorbance of the mixture at 540 nm was 0.18 in a 1 cm diameter cuvette. A standard solution (0.5 ml containing 4 mg of protein/ml) plus 4.5 ml of biuret reagent gave an absorbance of 0.12 in the same size cuvette.**

- a) Calculate the protein concentration in the undiluted unknown solution.**
- b) What is the composition of the blank here ?**

**A) Concentration of standard  $C_{st} = 4 \text{ mg/ml}$**

Thus  $C_{st} = 4 \text{ g/L}$

$$A_{\text{standard}} = a_s \times C \times l$$

$$0.12 = a_s \times 4 \times 1$$

$$\text{So } a_s = 0.12 / 4$$

$$= 0.03 \text{ (g/L)}^{-1} \text{ cm}^{-1}$$

**First**, find the value of  $a_s$  from the known values for standard solution

It is ( $a_s$ )  $\rightarrow$  because conc. of sample is in **mg/ml**

-To convert from: **mg/ml**  $\rightarrow$  **g/L** (divide by 1000)  
since the ratio is the same (1mg = 0.001 g and 1mL = 0.001L)

**Next**, apply the value of  $a_s$  on beer-lambert law to find the concentration of test

$$A_{\text{test}} = a_s \times C \times l$$

$\uparrow$  Dilution factor

$$0.18 = 0.03 \times C \times 1$$

$$\text{So } C_{\text{test}} = 0.18 / 0.03 = 6 \text{ g/L} = 6 \text{ mg/ml}$$

**The concentration of protein in the undiluted solution:**

$$Df = \text{total volume} / \text{aliquot volume} = (0.3 + 0.9) / 0.3 = 4$$

$$C_{\text{undiluted}} = 6 \times 4 = 24 \text{ mg/ml} .$$

**b) The blank should contain 4.5 ml of biuret and 0.5 ml of distilled water only.**

# EXAMPLE 4

## Solutions Containing Two Absorbing Substance

A solution containing NAD<sup>+</sup> and NADH had an absorbance of 0.311 in a 1 cm cuvette at 340 nm, and 1.2 at 260 nm. Calculate the concentration of the oxidized and reduced forms of the coenzyme in the solution.

**\*\*Both NAD<sup>+</sup> and NADH absorb at 260nm, but only NADH absorbs at 340nm.\*\***

$a_m$		
	260nm	340nm
NAD <sup>+</sup>	18000	0.0
NADH	15000	6220

**Note:** Absorbance at 340nm represents the absorbance of NADH only since NAD<sup>+</sup> does not absorb at that wavelength → So the concentration of NADH can be obtained.

\* At 340nm →  $A_{\text{NADH}} = a_m \times C \times l$

$$0.311 = 6220 \times C \times 1$$

So  $C_{\text{NADH}} = 0.311/6220 = 5 \times 10^{-5} \text{ M}$

\* At 280nm →  $A_{260\text{nm}} = A_{\text{NADH}} + A_{\text{NAD}^+}$  ( since both absorb at this wavelength )

$$A_{\text{NADH}} = a_m \times C \times l = 15000 \times 5 \times 10^{-5} \times 1 = 0.75$$

Thus  $A_{\text{NAD}^+} = A_{\text{total}} - A_{\text{NADH}} = 1.2 - 0.75 = 0.45$

Since  $A_{\text{NAD}^+} = a_m \times C_{\text{NAD}^+} \times l$

$$0.45 = 18000 \times C_{\text{NAD}^+} \times 1$$

$C_{\text{NAD}^+} = 0.45 / 18000 = 2.5 \times 10^{-5} \text{ M}$

# EXAMPLE 5

## Solutions Containing Two Absorbing Substance

Ten grams of butter were saponified , the non-saponifiable fraction was extracted into 25ml of chloroform . The absorbance of the chloroform solution in a 1cm cuvette was 0.53 at 328nm and 0.48 at 458nm . Calculate the carotene and vitamin A content of the butter .

$a_{1\%}$		
	328nm	458nm
Carotene	340	2200
Vitamin A	1550	0.0

$a_{1\%}$  = absorption coefficient when concentration expressed in 1g/100ml .



**Note:** The absorbance at 458nm represents the absorbance of Carotene only , thus its concentration can be obtained .

$$A_{458\text{nm}} = A_{\text{carotene}}$$

$$A_{458\text{nm}} = a_{1\%} \times C_{\text{carotene}} \times l = 2200 \times C_{\text{carotene}} \times 1$$

$$C_{\text{carotene}} = 0.48/2200 = 2.1 \times 10^{-4} \text{ g/100ml}$$

Thus the carotene content in the 25ml of chloroform extract is

$$2.1 \times 10^{-4} \text{ g} \text{ -----} \rightarrow 100\text{ml}$$

$$? \text{ -----} \rightarrow 25\text{ml}$$

$$\text{the carotene content in the 25ml of chloroform extract} = (25 \times 2.1 \times 10^{-4}) / 100$$

$$= 5.2 \times 10^{-5} \text{ g}$$

$$= 5.2 \times 10^{-2} \text{ mg .}$$

$$\text{The carotene content per gram of butter} = 0.052 / 10 = 5.2 \times 10^{-3} \text{ mg carotene / g of butter}$$

**Absorbance at 328nm is the absorbance of  $A_{\text{carotene}} + A_{\text{vitamin A}}$**

$$A_{\text{carotene}} = a_{1\%} \times C \times l = 340 \times 2.1 \times 10^{-4} \times 1 = 0.0714 .$$

$$A_{\text{vitamin A}} = A_{\text{Total}} - A_{\text{carotene}}$$

$$A_{\text{vitamin A}} = 0.53 - 0.0714 = 0.458$$

$$C_{\text{vitamin A}} = A / a_{1\%} \times 1 = 0.458/1550 = 2.9 \times 10^{-4} \text{ g/100ml}$$

Thus vitamin A content in the 25ml of chloroform extract is

$$2.9 \times 10^{-4} \text{ g} \text{ -----} \rightarrow 100\text{ml}$$

$$? \text{ -----} \rightarrow 25\text{ml}$$

$$\text{vitamin A content in the 25ml of chloroform extract} = (25 \times 2.9 \times 10^{-4}) / 100$$

$$= 7.25 \times 10^{-5} \text{ g}$$

$$= 0.073 \text{ mg}$$

$$\text{The vitamin A content per gram of butter} = 0.073/10 = 0.0073 \text{ mg / g of butter}$$

# COUPLED ASSAY

- Many compounds of biological importance do not have a distinct absorption maximum  $\lambda_{\max}$  .
- Their concentration can be determined if they can be linked to or coupled with a reaction that fulfills the following condition ..

*If they promote the formation of another compound that has a characteristic absorption peak.*

- In coupled assay reactions the product of the first reaction is the substrate of the following reaction ;



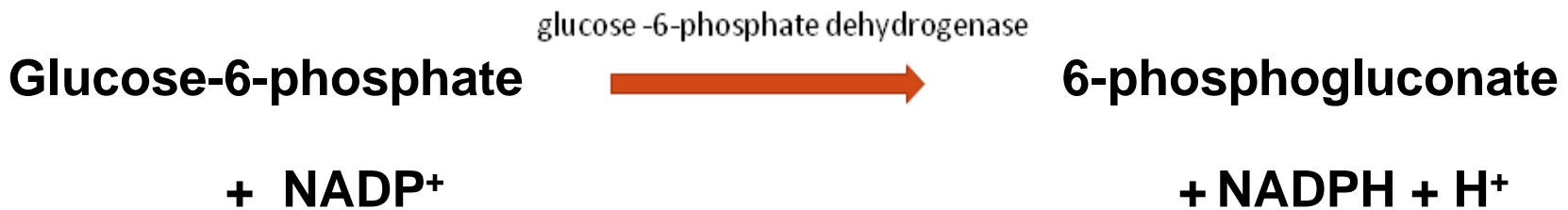
A = substance under study that does not have a distinct  $\lambda_{\max}$

F = has a distinct  $\lambda_{\max}$

*→ Thus A can be estimated by measuring the absorbance of F*

# EXAMPLE

To 2.0 ml of a glucose solution; 1.0 ml of a solution containing excess ATP, NADP<sup>+</sup>, MgCl<sub>2</sub>, hexokinase and Glucose -6-phosphate dehydrogenase was added. Calculate the concentration of glucose in the original solution. Absorbance of final solution at 340 nm increased to 0.91,  $a_m = 6220$ ,  $L = 1$  cm.



## Notes:

\***Glucose** has no absorption at 340 nm, but **NADPH** does!

\*From the reaction: 1 mole of glucose produces 1 mole of NADPH, thus each number of NADPH moles produced originates from every mole of glucose in the original solution.

Absorbance at 340 nm is the absorbance of NADPH = 0.91

$$A = a_m \times C \times L \rightarrow 0.91 = 6220 \times C \times 1$$

$$C_{\text{NADPH}} = 0.91 / 6220 = 1.46 \times 10^{-4} \text{ M}$$

Thus there is  $1.46 \times 10^{-4} \text{ M}$  of glucose present in the *test* solution

$$\begin{aligned} \text{The glucose concentration in the } \textit{original} \text{ solution} &= 1.46 \times 10^{-4} \times (3/2) \\ &= 2.2 \times 10^{-4} \text{ M} \end{aligned}$$



Dilution factor