# Agarose Gel Electrophoresis of DNA

By: Sahar alSubaie

principle :

Agarose Gel Electrophoresis uses electrical field to separate macromolecules (DNA and Protein) that differ in <u>size</u>, <u>charge</u> and <u>configuration</u>.

#### DNA :

DNA molecules have –ve charges.
 They differ in size (bp).

# Factors affect electrophoresis

# Molecule's charge

# DNA (-ve charged molecule) moves from cathod (-ve) to Anode( +ve) pole



# Molecule's size

#### The lighter the molecule the faster it is .



# The Agarose concentration:

- The higher the concentration the smaller the size of molecule that can pass the matrix.
- The higher the concentration of gel the slower the movement of particles.
  - 2% is the concentration of choice for separation of most of DNA fragments.



The gel matrix EM. Image

# Conformation of DNA.

The liner DNA is slower than supercoiled molecule .



#### **Applied current**

In general ,the higher the current ampere and Voltage the faster the net movement of molecules

# **Direction of electrical field**

- For DNA separation the direction is from cathode to anode
- So make sure to put your sample at the right direction or no movement will result.
- Be carful to choose the right electrical connection of the device too.

# electrophoresis buffer

As it provides the ions which transfer the electrical filed within the fluid .

Also they modify the net charges of the molecules

# Intercalating dyes

Ethidium Bromide binds (intercalation between the base pairs) to the DNA and decrease its mobility.

The larger the DNA molecule the more Ethidium Bromide binds to it.



#### Equipment of agarose gel electrophoresis

Electrophoresis chamber . Power supply. Gel casting trays. □ Sample combs to form sample wells. Running buffer. Loading buffer. **D**. Ethedium bromide. (mutagen) Transilluminator.



Gel casting tray & combs

### **Running buffers:**

Choosing depends on the size of DNA.
 <u>TAE buffer (Tris Acetate EDTA)</u> :the most common used buffer.
 <u>TBE buffer (Tris Borate/EDTA)</u> is often used for smaller DNA fragment (i.e less than 500bp).

# Loading buffer:

This buffer mixed with the sample, it gives the samples :

- ✓ color
- ✓ density
- ✓ makes it easy loading in the wells.

It consists of bromophenol blue( the tracing dye), sucrose and water.

# Markers (DNA ladder):

#### Different kinds of markers

- most of them are bacteriophage DNA cut with restriction enzymes
- Storing markers ready mixed with loading buffer at 4C.
- Choosing marker with a good resolution for fragment size in the experiment.



DNA size marker

# Visualization:

Transiluminator (an ultraviolet light box) is used to visualize :✓ Ethidium bromide- stained DNA in gels.

✓ Ethidium bromide-free techniques are available also.

# **Purposes**:

To look at the DNA
To quantify it
To isolate a particular band.

## **Applications**:

- Estimation DNA quantity and quality
- Using a DNA ladder to approximate the size of DNA molecules.
- Analysis of PCR products.
- Separation of restriction enzyme digested DNA prior to Southern Blot transfer.
- Quantity is assessed using lambda DNA ladder.
- Quality of DNA is assessed by observing the absence of streaking or fragments (or contaminating DNA bands).

### **Procedure:**

- Prepare a 1% agarose solution, measure 3 g agarose into a flask and add 150 ml 1X buffer. Microwave until agarose is dissolved and solution is clear.
- Allow solution to cool to about 55°C before pouring. Add 10 μl of Ethidium bromide or alternative dye
- Prepare gel tray and place comb in gel tray.
- Pour 50°C gel solution into tray and allow gel to solidify about 20 minutes at room temperature.
- To run, gently remove the comb, place tray in electrophoresis chamber, and cover (just until wells are submerged) with electrophoresis buffer (the same buffer used to prepare the agarose).
- To prepare samples for electrophoresis, add 1 μl of 6x gel loading dye for every 5 μl of DNA solution. Mix well & load. Don't forget to load the DNA size marker.
- Electrophorese at 50-150 volts until dye markers have migrated an appropriate distance, depending on the size of DNA to be visualized.
  - Visualize DNA under the UV transilluminator.

# **References:**

- OpenWetWare contributors, "20.109(F07): Agarose gel electrophoresis," OpenWetWare, , http://openwetware.org/index.php?title=20.109%28F07%29:\_Agarose\_gel\_electrophoresis&oldid=153043 (accessed August 29, 2016).
- Tirabassi, R. (2016) How to identify Supercoils, Nicks and circles in Plasmid Preps. Available at: http://bitesizebio.com/13524/how-to-identify-supercoils-nicks-and-circles-in-plasmid-preps/ (Accessed: 29 August 2016).
- Sigmon, J. and Larcom, L. (1996) 'The effect of ethidium bromide on mobility of DNA fragments in agarose gel electrophoresis', Electrophoresis., 17(10), pp. 1524–7.
- Inc, T.F.S. (2015) Nucleic acid Stains—Section 8.1. Available at: https://www.thermofisher.com/sa/en/home/references/molecular-probes-the-handbook/nucleic-aciddetection-and-genomics-technology/nucleic-acid-stains.html (Accessed: 29 August 2016).
- Sambrook and Russell (2001). Molecular Cloning: A Laboratory Manual (3rd ed.). Cold Spring Harbor Laboratory Press. (Sambrook and Russell cites the paper:Glasel J. (1995). "Validity of nucleic acid purities monitored by 260nm/280nm absorbance ratios". BioTechniques 18: 62–63.)
- Clark, W. and K. Christopher. 2000. An introduction to DNA: Spectrophotometry, degradation, and the "Frankengel' experiment. Pages 81-99, in Tested studies for laboratory teaching, Volume 22 (S. J. Karcher, Editor). Proceedings of the 22nd Workshop/Conference of the Association for Biology Laboratory Education (ABLE), 489 pages.
- The Analysis of DNA or RNA using Its Wavelengths: 230 nm, 260 nm, 280 nm". Bioteachnology.com. 2010-01-13. Retrieved 2010-03-12.