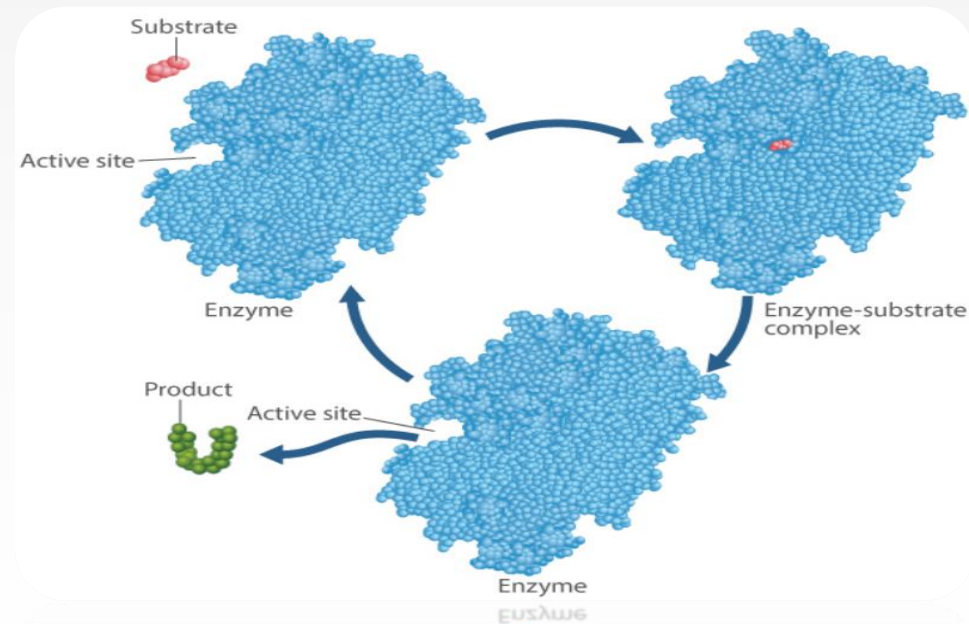
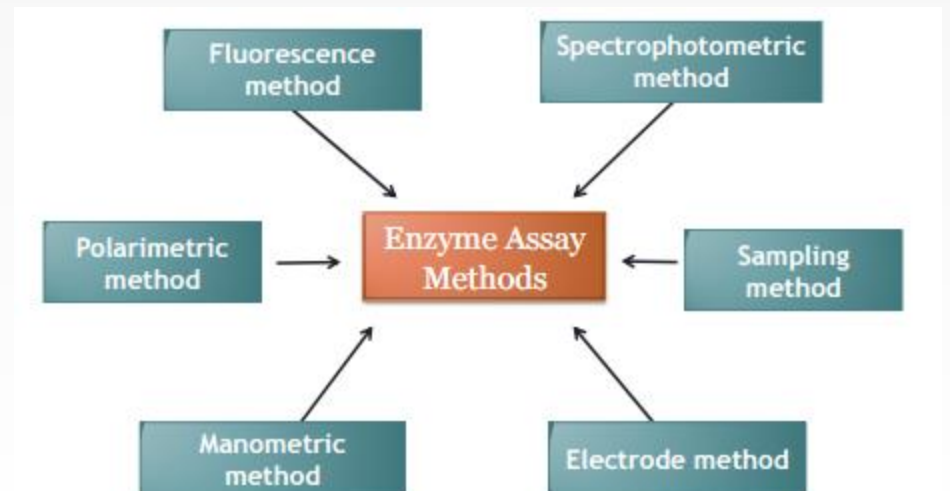


# Method of Enzyme Assay



## Objective:

- To study the different methods for determining enzyme activity.
- Use these method in diagnosis of certain diseases



## How to follow a reaction?

*Enzyme assays*: Are laboratory methods for measuring **enzymatic activity**.

- Enzyme assays measure either the **consumption of substrate** or **production of product** over time.



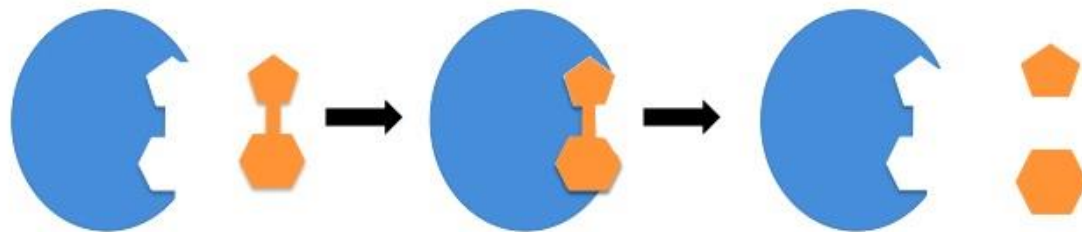
- **Different enzymes require different estimation methods** depending on the type of reaction catalyzed, the nature of S and P or coenzyme.

# Methods of quantitatively following enzyme reactions

## How to follow an enzymatic reaction?

First you must have complete knowledge about the reaction itself,

- Does the substrate or product has the ability to absorb light, fluorescence, any production of gases, production of  $H^+$ ?
- After that you can use this properties to detect the reaction



Enzyme + Substrate

Enzyme-Substrate  
Complex

Enzyme + Products

# Methods of quantitatively following enzyme reactions

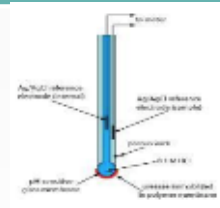
**Fluorescence methods**  
Using fluorometer



**Manometric methods**  
Using manometer.



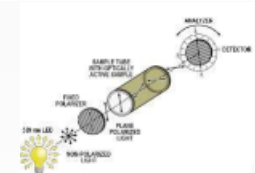
**Electrode Methods**  
Using a pH meter



**Spectrophotometric methods**

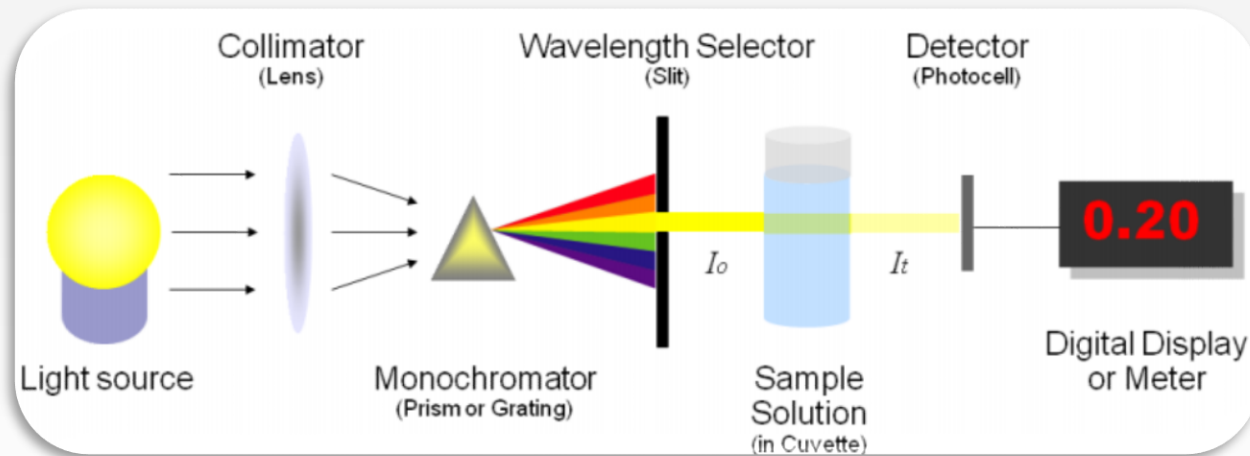


**Polarimetric Method**  
polarimeter



# Spectrophotometric method Principle

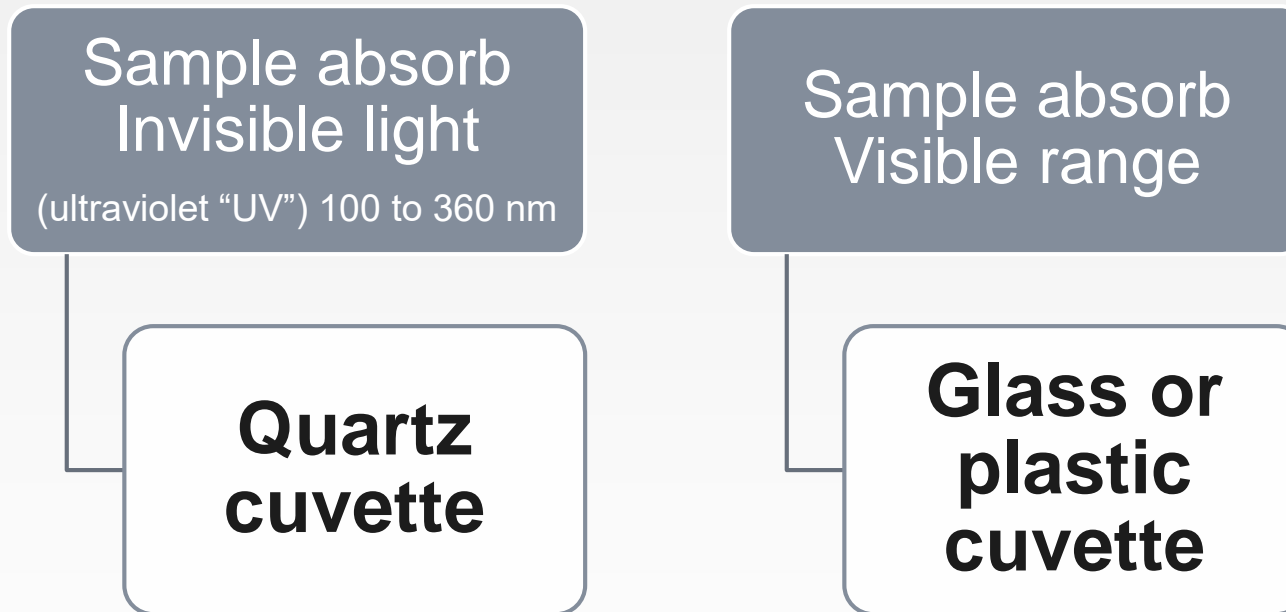
In spectrophotometric assays, you follow the course of the reaction **by measuring a change in how much light the assay solution absorbs.**



**What is blank solution?**

It is a solution that contains everything except the compound to be measured.

# Spectrophotometric methods



If the light is in the visible region you can actually see a change in the color of the assay, these are called [**colorimetric assays** ]

# Examples of using Spectrophotometric methods

1- cases in which product absorb but not the substrate.

e.g.

**Fumarate**

**Fumarate hydratase**

**malate**

Light

2- the Co-enzyme undergoes change in absorption upon reduction or oxidation

Oxidized form

**NAD**

**NADP**

Reduced form

**NADH**

**NADPH**

UV. Light  
340 nm

If **reduced form** was **product**: **increase** the absorbance / min

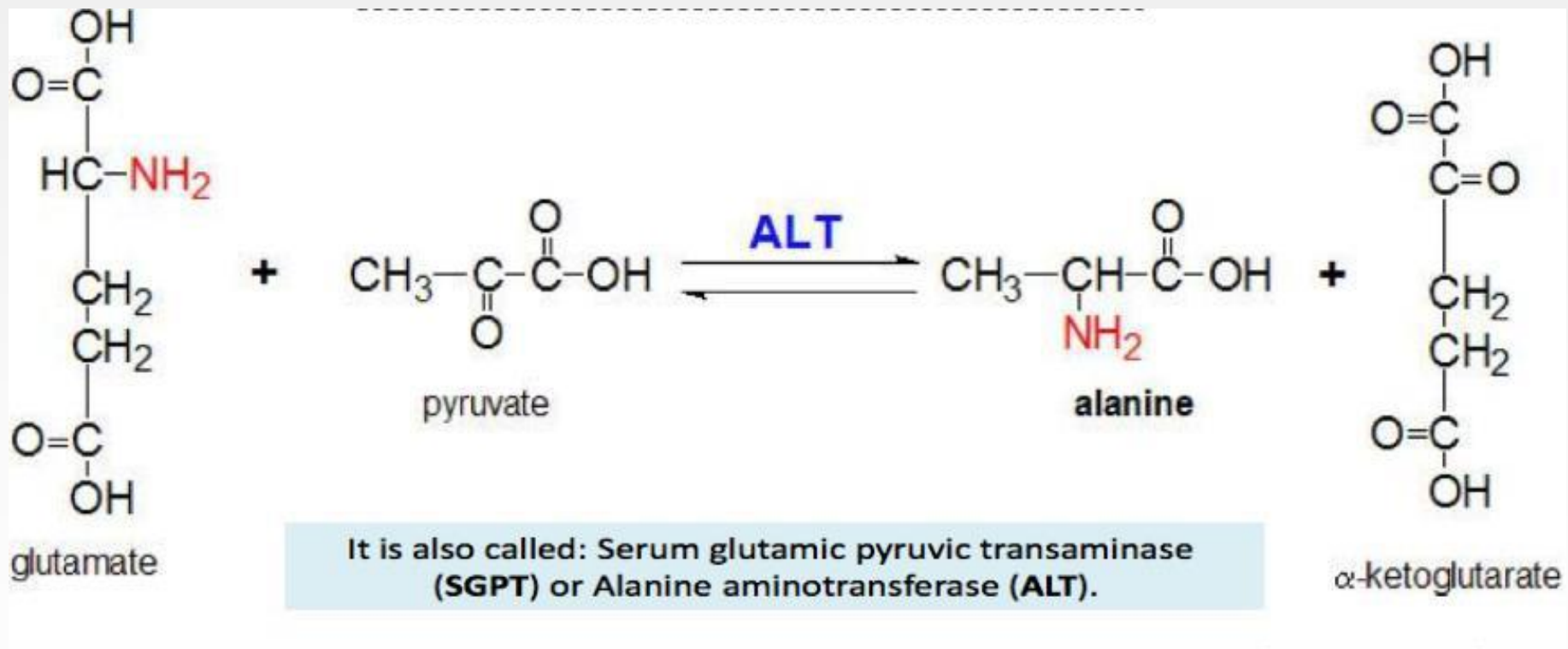
If reduced form was **substrate** : **decrease** the absorbance / min



## Two types of Enzyme assays:

- **Continuous assays**, where the assay gives a **continuous reading** of activity.
- **Discontinuous (Endpoint) assays**, Where the reaction is **stopped** and then the concentration of substrates/products determined.

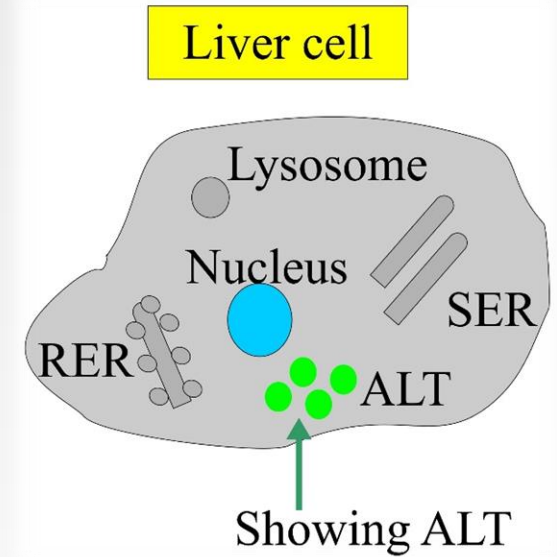
## If Substrate and Product can not absorb light? What is the solution?



# Alanine transaminase (ALT) in serum

Continues assay

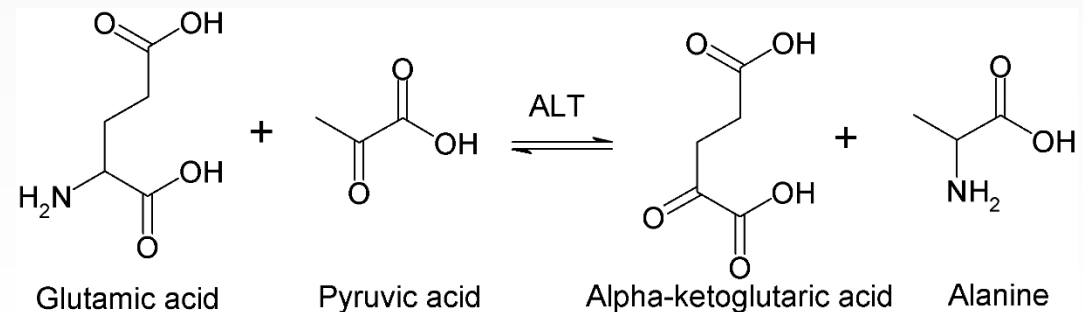
Discontinues assay



# Alanine transaminase (ALT)

- ALT is an enzyme that catalyzes a type of reaction (transamination) between an amino acid and  $\alpha$ -keto acid.
- It is important in the production of various amino acids.

- Transfer of amino group of an amino acid to  $\alpha$ -keto acid resulting in formation of new amino acid and new keto acid.

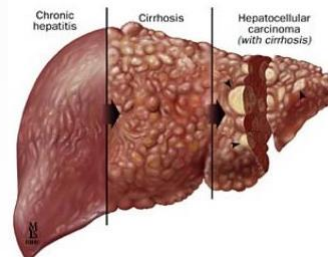


# ALT diagnostic importance



- ALT is found in serum (**at low level**) but is most commonly found in liver.
- Thus , an **elevated** level of ALT is a sensitive index of *acute hepatocellular injury*.
- Elevated serum ALT levels are found in hepatitis, cirrhosis , and obstructive jaundice.

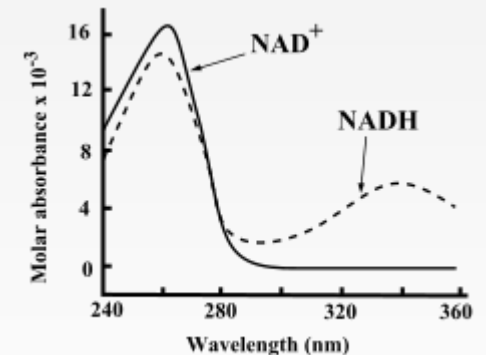
• **NORMAL RANGE OF ALT:** ( up to 42 ) U/L → males ( up to 32 ) U/L → females



# 1-Continuous Assay

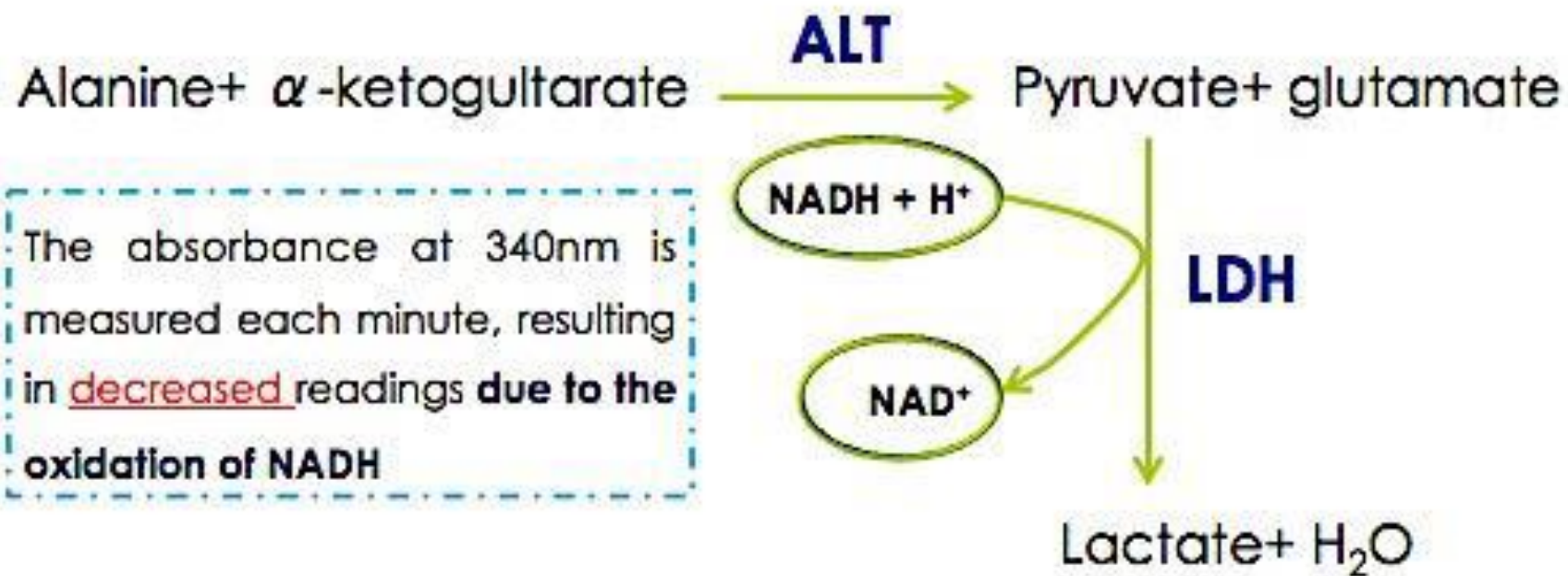
- The reading will be continuous (1 min for 3 min).
- Because neither the S nor P can absorb light, the following can be done:  
**First we will add the enzyme to the Substrate**

1. Alanine +  $\alpha$ -ketoglutarate  $\rightarrow$  Pyruvate + glutamate
2. Another enzyme (LDH) and NADH+H<sup>+</sup> will be added:
3. Pyruvate + NADH+H<sup>+</sup>  $\rightarrow$  L-Lactate+ NAD<sup>+</sup> + H<sub>2</sub>O



The absorbance at 340nm is measured each minute without stopping the reaction, resulting in **decreased** readings **due to the oxidation of NADH**.

## Principle



## Method

Pipette into clean and dry test tubes:

| ALT Reagent   | 3ml                   |
|---|-----------------------|
| Pre-warm at 37°C for 3 minutes and add  |                       |
| Serum Sample  | 0.2 ml = (    μl) ??? |
| Mix and incubated at 37 °C for 1 minute, then read absorbance ( at 340 nm against distilled water ) every minute for 3 minutes) and determine $\Delta A/\text{min}$ |                       |

**Choose the following on the spectrophotometer:**

**2) Applications → 2) Simple Kinetics → wave length (340 nm) → Seconds → Duration (180 sec = 3 min) → Intervals (60 sec= 1 min) → Print Data Table (off) → Press start (2 times)**



## Results

| Time  | Absorbance 340nm | $\Delta A/\text{min} = ((A1-A2)+(A2-A3))/2$ |
|-------|------------------|---|
| 1 min | A1               |   |
| 2 min | A2               |   |
| 3 min | A3               |   |

## Calculations:

ALT Activity ( U/L) =  $\Delta A/\text{min} \times 1768$

ALT Activity (U/L) = **Calculations**

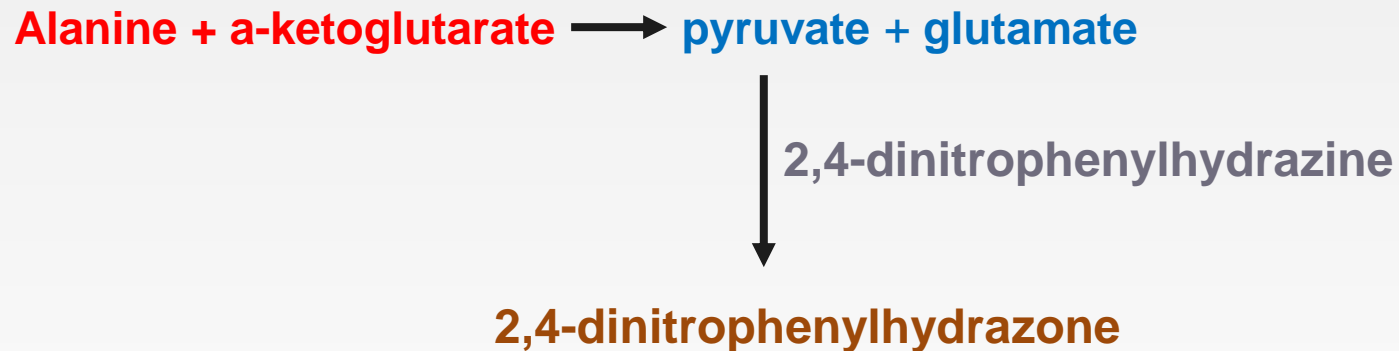
# Discontinuous Assay

- In this method **ALT** catalyzes the following reaction



- ALT is assayed by following formation of **pyruvate**.
- The addition of acidic 2,4-dinitrophenylhydrazine (DNPH) lead to the formation of 2,4-dinitrophenylhydrazone, then NaOH will be added So that it may be measured at 546nm.

# Reaction-discontinuous



- This assay as an example of **colorimetric\ endpoint assay**
- **Why colorimetric?** Because it will give a brown color
- Normal Range: 10-40 units per liter (U/L)

## Method:

|  | BLANK  | SAMPLE |
|--|--------|--------|
| ALT Reagent  | 0.5 ml | 0.5 ml |
| Pre-warm at 37 °C for <u>5 minutes</u> and add:  |        |        |
| Distilled Water  | 0.1 ml | -      |
| Serum Sample   | -      | 0.1 ml |
| Mix, and incubate at 37 °C for exactly <u>30 minutes</u> , and add:  |        |        |
| Color Reagent<br>(DNPH)  | 0.5 ml | 0.5 ml |
| Mix, and return at 37 °C for exactly <u>10 minutes</u> , then add:   |        |        |
| Color Developer<br>(NaOH)  | 5.0 ml | 5.0 ml |
| Mix, and return to 37 °C for exactly <u>5 minutes</u> . Read absorbance of all tubes at 546nm against blank. |        |        |

## Precautions



- COLOR REAGENT contains 1 N Hydrochloric acid which **causes burns**.
- COLOR DEVELOPER contains 0.5 N Sodium hydroxide which is **corrosive**.

In case of contact, flush affected area with large amounts of water. Seek medical attention.

## Results:

| Absorbance at 546 nm | ALT Activity (U/L) |
|----------------------|--------------------|
| 0.025                | 2.5                |
| 0.050                | 5.5                |
| 0.075                | 9                  |
| 0.100                | 12                 |
| 0.125                | 17                 |
| 0.150                | 21                 |
| 0.175                | 25                 |
| 0.2                  | 30                 |
| 0.225                | 35                 |
| 0.250                | 41                 |

- The data shown in the table is used to convert absorbance at 546 nm into enzymatic activity in U/L of serum.
- Draw graph using the data in table with absorbance on the Y- axis and enzymatic activity in U/L on the X-axis.

**Note:** Don't forget title of the graph "Standard Curve" and the x- axis and y- axis with their units

**-Absorbance at 546 nm = .....**

**-ALT (SGPT) activity (from graph)= .....**

## Discussion:

- Mention the diagnostic importance of ALT
- Explain the difference in the principle of each ALT assay.
- Compare your result with ALT normal range [in males], and diagnose the patient's state (what disease could the patient have or not).