

Quantitative estimation of proteins by Bradford methods

Materials:

Chemical

Prepared crude extract, 40% pellet, dialyzed sample, BSA standard solution (2 g/l), Bradford reagent, distal water.

Preparation of solutions

1) Bradford reagent

- Dissolve 100 mg of Coomassie brilliant-G250 in 50 ml of 95% ethanol, add 100 ml of 85% w/v phosphoric acid and then complete the volume to 1 L by adding dis. H₂O.
- After the dye has completely dissolved, filter through Whatman#1 filter paper just before use.
- Filtration may have to be repeated to get rid of all blue components.
- Keep in dark bottle.

Equipment and Glassware

Micropipette, tips, plastic cuvettes, spectrophotometer.

Protocol:

1. In plastic cuvettes ad the following:

	BSA standard solution (5 g/l) volume (μ l)	Distal water (μ l)	Sample (μ l)	Bradford reagent (μ l)
Blank	-	100	-	1000
A	5	95	-	
B	10	90	-	
C	20	80	-	
D	40	60	-	
E	60	40	-	
F	80	20	-	
G	100	-	-	
Animal crude extract (D1)	-	90	10	
Animal crude extract (D2)	-	80	20	
Plant crude extract (D1)	-	90	10	
Plant crude extract (D2)	-	80	20	
40% pellet	-	80	20	
Dialyzed sample	-	80	20	

2. Mix the content of each tube.
3. Incubate for **15 min** at room temperature.
4. Read the absorbance at **595 nm** against blank.

5. Determine the protein contents from BSA standard curve.

Results:

Test tube	Protein concentration ($\mu\text{g/ml}$) [X- axis]	Absorbance at 595 nm [Y- axis]
Blank		
A		
B		
C		
D		
E		
F		
G		
Animal crude extract (D1)	_____	
Animal crude extract (D2)	_____	
Plant crude extract (D1)	_____	
Plant crude extract (D2)	_____	
40% pellet	_____	
Dialyzed sample	_____	