



Biochemistry of Proteins BCH 303 [Practical]

Lab (4) Protein extraction from animal and plant source

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Cell lysis and disruption

- **Cell lysis** is the first step in cell fractionation, organelle isolation and protein extraction and purification.
- It opens the door to a myriad of proteomics research methods.

- **Many techniques** have been developed and used to obtain the best possible yield and purity for:
 1. Different species of organisms
 2. Sample types (cells or tissue)
 3. Target molecule or subcellular structure

- **Cell disruption methods can be categorized into:**
 - a. Mechanical methods
 - b. Non-mechanical methods

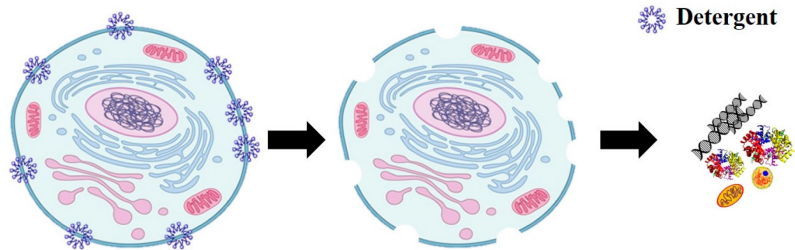
Cell disruption methods

Mechanical methods:

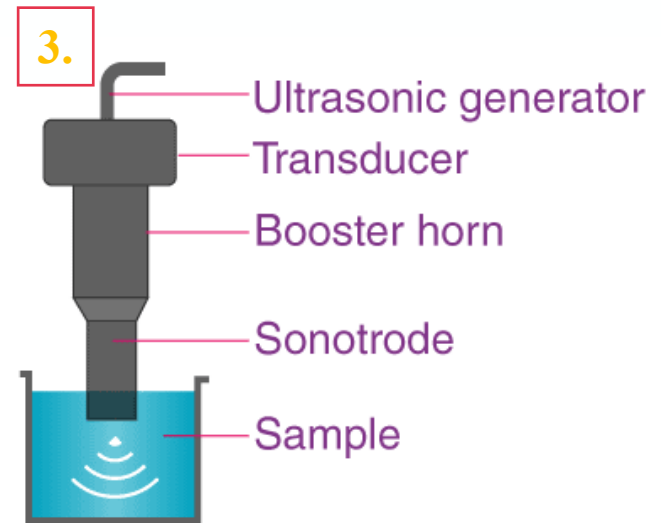
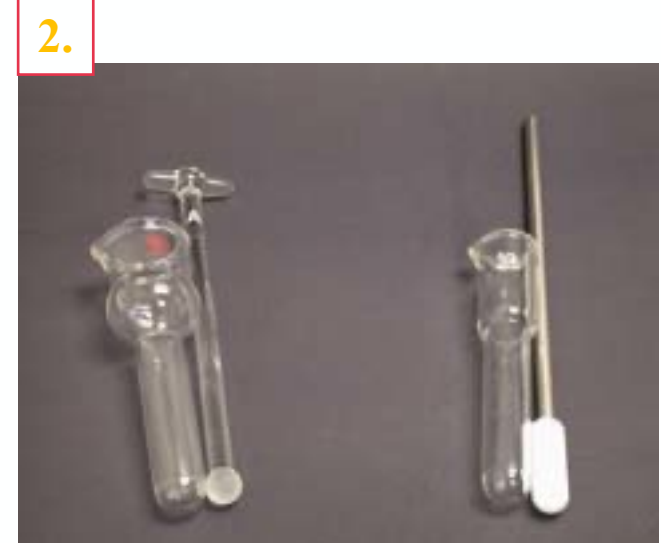
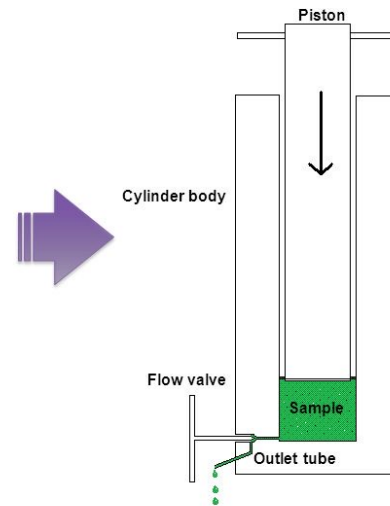
1. French Press
2. Mortar and pestle
3. Sonication

Non-mechanical methods:

1. Detergent-based cell lysis




4. Detergent reacts with cell membrane
- Detergent destroys the cell membrane
- Intracellular components are released



Protein extraction/isolation

- There is no universal protocol for protein sample preparation. (*Why?*)
- **Sample preparation protocols must take into account several factors:**
 1. The **source** of the specimen or sample type
 2. **Chemical** and **structural** heterogeneity of proteins
 3. The cellular or subcellular **location** of the protein of interest
 4. The required protein **yield**.
 5. Proposed downstream application.

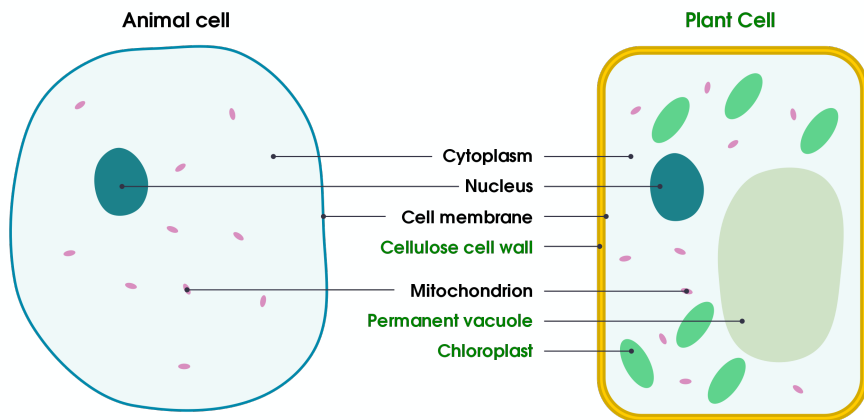
 PAUSE AND THINK → How cellular location of the protein will affect the isolation protocol?

Practical part 

Principle

- The **initial step** of any purification procedure must, of course, be to disrupt the starting tissue to release proteins from within the cell.
- The **means of disrupting** the tissue will depend on the cell type.
- **In animal cells**, the plasma membrane is the only barrier separating cell contents from the environment.
- **In plants and bacteria** the plasma membrane is also surrounded by a rigid cell wall.
 - a. Plant cell walls consist of multiple layers of cellulose.
 - b. These types of extracellular barriers confer shape and rigidity to the cells.
 - c. Plant cell walls are particularly strong, making them very difficult to disrupt mechanically or chemically.

Whereas the *lack* of an extracellular wall in animal cells → makes them relatively easy to lyse.



Principle

- Cell disrupting will be achieved using both **mechanical** and **non-mechanical** methods.
- **In non-mechanical method:**
 - Normally extraction buffers are at an **ionic strength (0.1–0.2 M)** and **pH (7.0–8.0)** that is considered to be compatible with that found inside the cell.
 - **Tris** or **phosphate buffers** are most commonly used.
- **In the mechanical method:**
 - Both animal and plant cells are susceptible to **shear forces using blenders**.
 - The tissue is cut into small pieces and blended, in the presence of buffer to disrupt the tissue, and then centrifuged to remove debris.
- After extraction, **protein concentration determination** is a routine requirement during protein purification, which can be achieved by different method. (*coming labs*)

Experiment 1. Protease inhibitor extraction from plant source

Aim:

- To prepare crude extract from plant source.

Method:

1. Weight 12 g of the sample and place it in the blender with 200 ml of the extraction buffer (phosphate buffer 0.1 M, pH 7.0)
2. Incubate the homogenate at room temperature on a rotary shaker for 30 min at 150 rpm.
3. Filter the slurry through cheesecloth and then transfer to centrifuge tube.
4. Centrifuge the filtrate at 10,000 rpm for 10 min at 4 °C for the removal of any cell debris that remained in the preparation.
5. Measure the volume of the supernatant.

Experiment 2. Lactate dehydrogenase extraction from animal source

Aim:

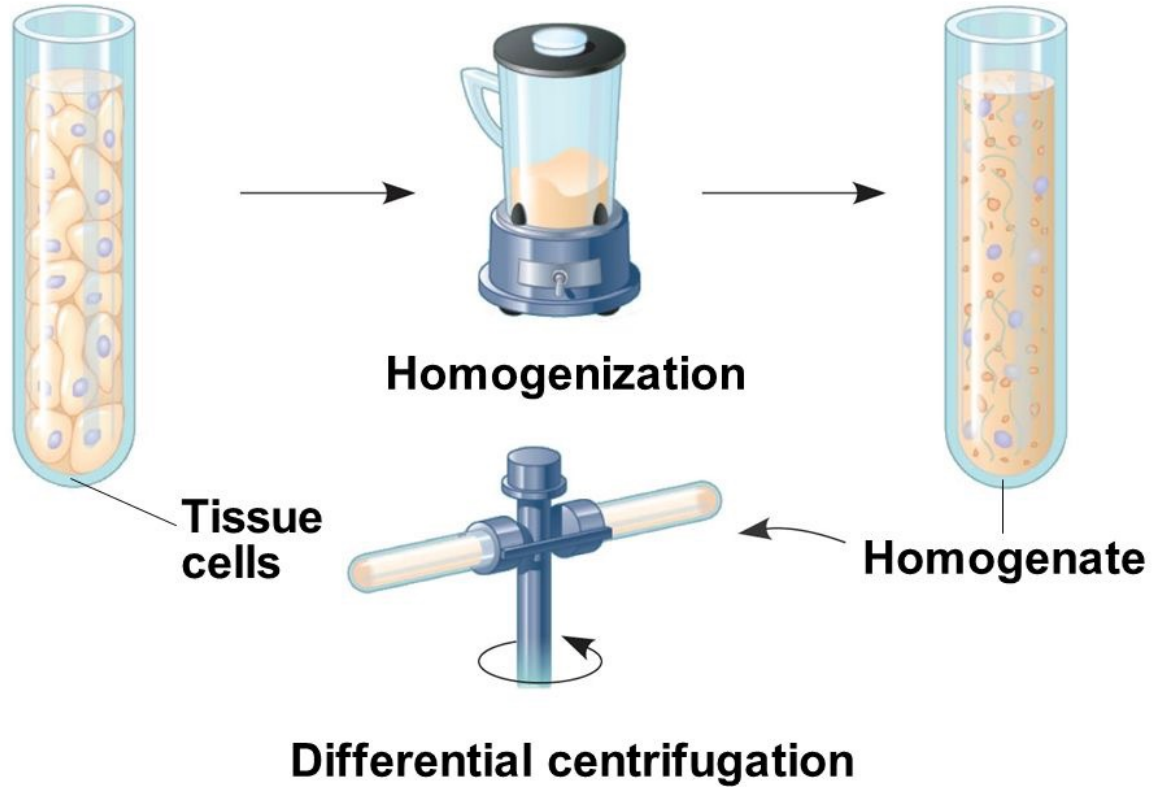
- To prepare crude extract from animal source.

Method:

1. Cut ~7.5 g of muscle tissue from the tissue source. *Record the exact weight of tissue used.*
2. Cut the tissue into small pieces. *Discard the connective tissue and fat.*
3. Add 38 ml of cold extraction buffer (0.1 M Tris-HCl, pH 7.4) in a blender with the sample. *Note: (20% weight/volume).*
4. Transfer the homogenized tissue/buffer mixture into centrifuge tubes.
5. Centrifuge your homogenate for 10 minutes at 7000 rpm.
6. Measure the volume of the supernatant.

Shear forces using blenders

LE 6-5a



Homework

- There are different mechanical methods (other than using blenders) for cell disruption. Search for one method and describe it briefly.