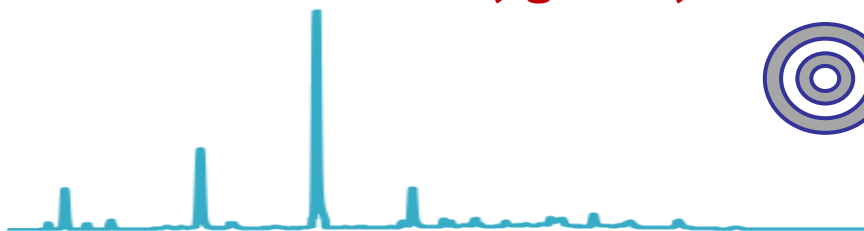
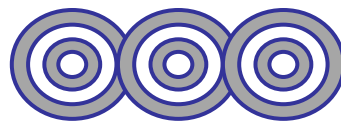


Chem 651

Advanced Studies in Instrumental Analysis

Miniaturization of Liquid Chromatography

Towards sensitive, green, and economic chromatographic techniques



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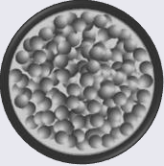
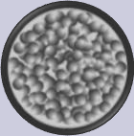


كرسي أبحاث
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Research Chair



Liquid Chromatography Scales

The column is considered the heart of the **HPLC** & **GC** techniques. The separation of the sample components is achieved when those components pass through the column.

The move to smaller i.d. columns

	HPLC columns were originally 4.6 mm i.d. operated at 1.0–2.0 mL/min
	3.0 mm i.d. columns introduced as a means to save solvents 50% solvent savings going from a 4.6x100 mm at 1.5 mL/min to a 3.0x100 mm at 0.75 mL/min
	Short columns with 2.1 mm i.d. and sub-2 μ m stationary phases particle size introduced for use with UHPLC
	In 1988, K. Karlsson & M. Novotny published a paper “separation efficiency of slurry packed liquid chromatography microcolumns with very small inner diameters”, they reported high efficiencies with micro-LC columns that had the i.d. of 44 μ m

Since that time great efforts have been made to miniaturize **LC** instrumentation by carrying out theoretical, technological, and methodological studies.

Definitions for the miniaturized chromatographic system

LC technique	Column i.d.	Typical flow rate	Injection volume	Relative sensitivity
Preparative	> 25 mm	> 30 mL/min	--	--
Semi-preparative	5–10 mm	5–20 mL/min	--	--
Conventional	3.2–4.6 mm	0.5–2.0 mL/min	100 µL	1
Narrowbore	1.5–3.2 mm	100–500 µL/min	19 µL	5
Microbore	0.5–1.5 mm	10–100 µL/min	4.7 µL	21
Micro LC capillary	150–500 µm	1–10 µL/min	490 nL	235
Nano LC capillary	10–150 µm	10–1000 nL/min	12 nL	3800

LC technique	Column i.d.	Typical flow rate	Injection volume
Preparative	> 5 mm	> 5 mL/min	---
Conventional	2.1–4.6 mm	0.5–2.5 mL/min	5–25 µL
Micro LC	0.5–2.1 mm	10–500 µL/min	0.5–5 µL
Capillary LC	100–500 µm	0.5–10 µL/min	10–500 nL
Nano LC	10–100 µm	10–500 nL/min	< 10 nL

Motivation Behind Downscales LC



Advantages of using down scales LC

Improving detection sensitivity

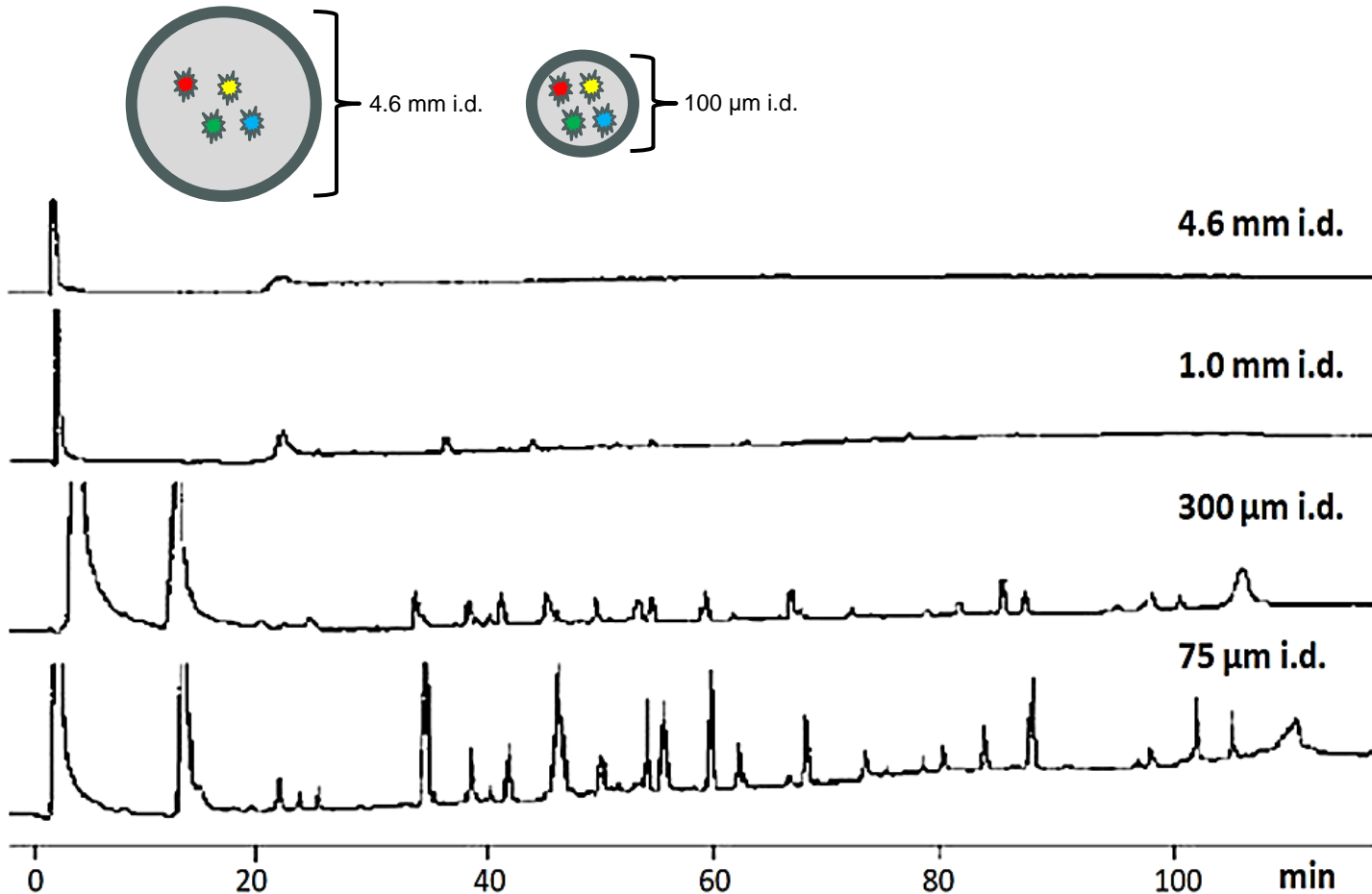
Easier coupling to MS interfaces

Lower solvent & sample consumption

Reduce analytical costs and wastes

Less subject to the effects of viscous heating

Improving detection sensitivity



A sensitivity comparison of different column i.d. for digested myoglobin.

↓ Column i.d. → ↓ Sample dilution & diffusion → ↑ Sensitivity

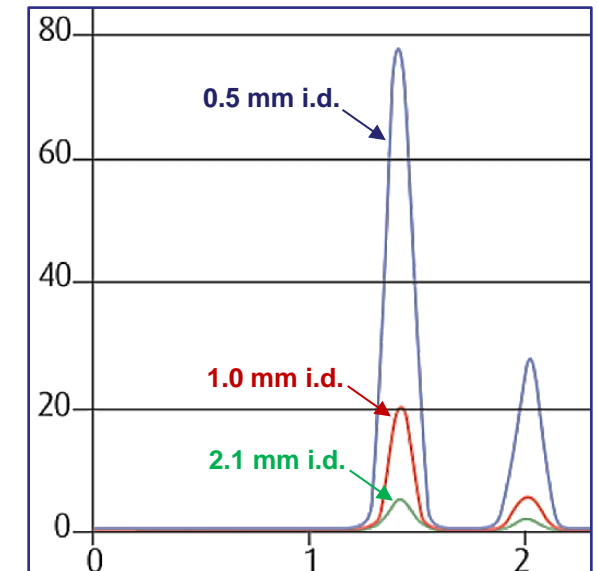
↑ Sensitivity → ↓ Detection limits

Down scale factor:

$$f = \frac{(i.d.thicker)^2}{(i.d.thinner)^2}$$

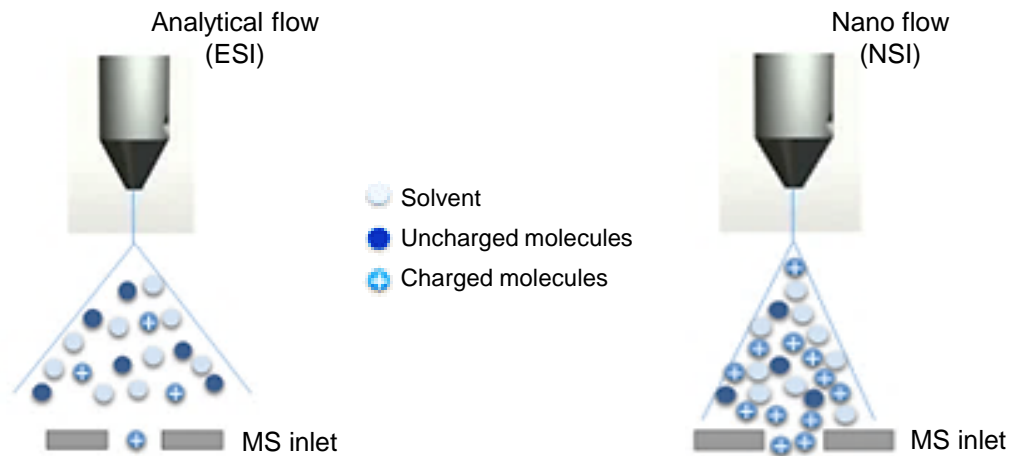
Example: (4.6 mm vs. 100 µm)

$$f = 4600^2 / 100^2 = 2100$$



Easier coupling to mass spectrometer interfaces

Better flow rate compatibility with MS detectors.



analytical vs. low flow ionization

The smaller volumetric flow rate results in smaller droplet sizes, that are easily evaporated, which increase the ionization efficiency and improve detection sensitivity.

Less subject to the effects of viscous heating

Column i.d.	Flow rate	Power
4.6 mm	2.0 mL/min	24 W
2.0 mm	0.38 mL/min	4.5 W
1.0 mm	95 µL/min	1.1 W
500 µm	24 µL/min	280 mW
250 µm	5.9 µL/min	71 mW
100 µm	940 nL/min	11 mW
50 µm	240 nL/min	2.9 mW

Considerations for reducing viscous friction.

$$\text{Power} = F \Delta P$$

1 W	= 14.33 calories/min
	= 7.17 calories/mL

Power represents the enough power to boil mobile phase before it exits the column.

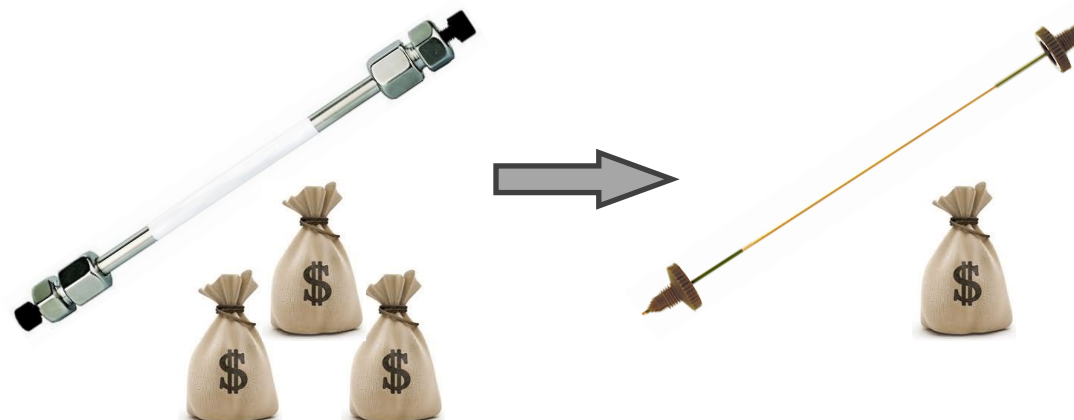
Lower solvent and sample consumption

Lower mobile phase solvents, samples, and stationary phase materials consumption.

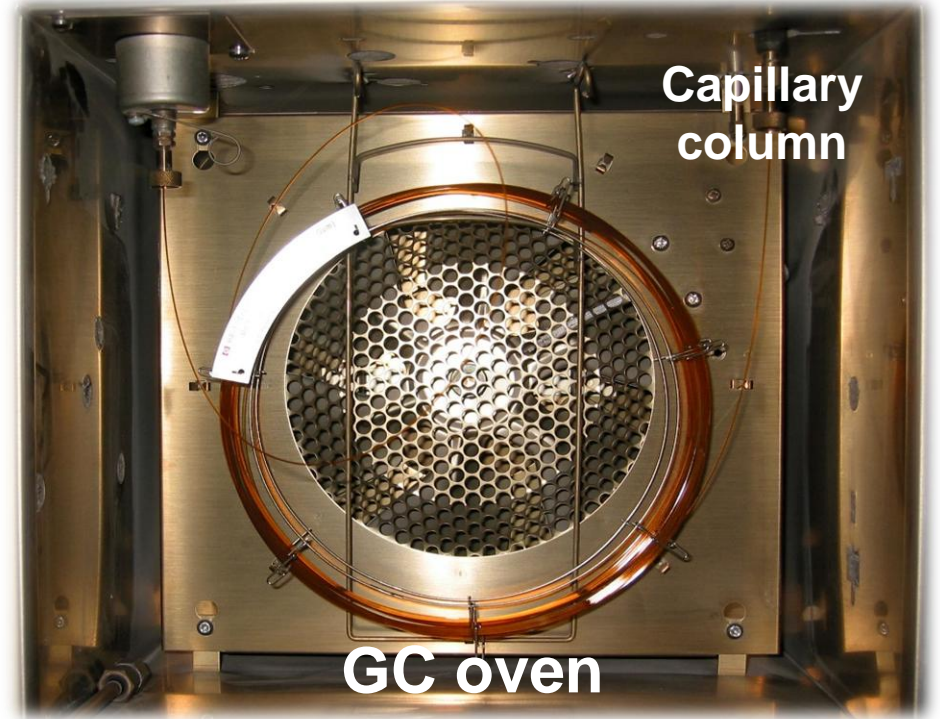
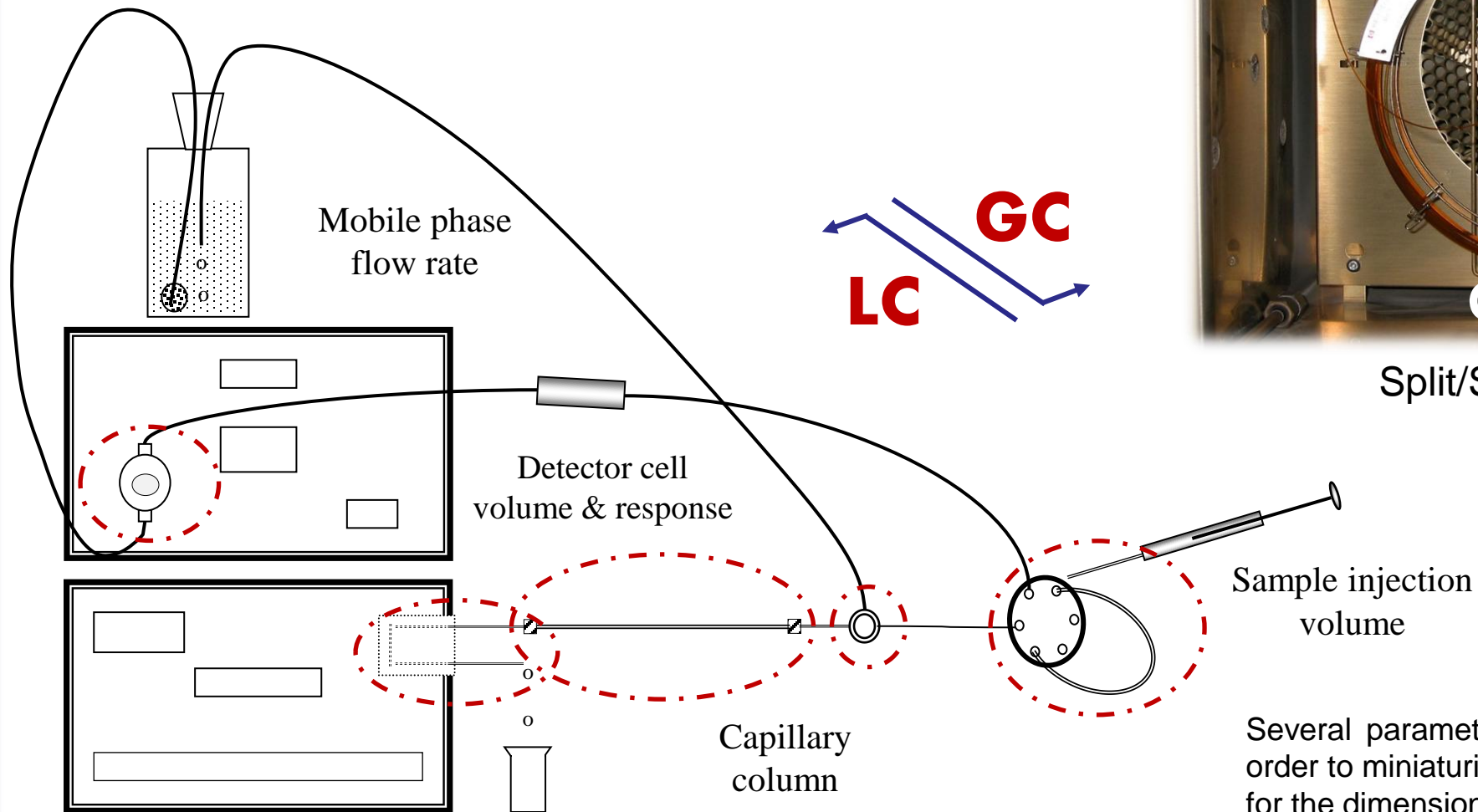
4.6 mm column i.d.	15 cm column length	100 μ m column i.d.
1.0 mL/min 60 mL/h	Solvents (mobile phase) Flow rate	1.0 μ L/min 60 μ L/h (~1000 times lower)
10 μ L	Samples (injection volume)	4.0 nL (~2500 times lower)
2.5 mL	Materials (stationary phase) Empty column volume	1.18 μ L (~2100 times lower)

Good for the expensive samples and deuterated solvent in the coupling of **LC** to **NMR**.

Reduce analytical costs and wastes



Miniaturized LC System Challenges

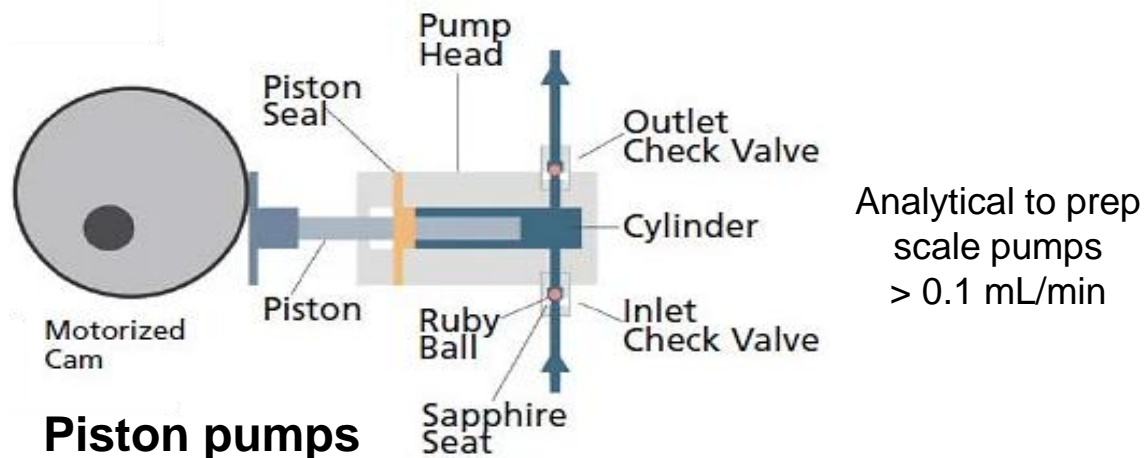


Split/Splitless injection

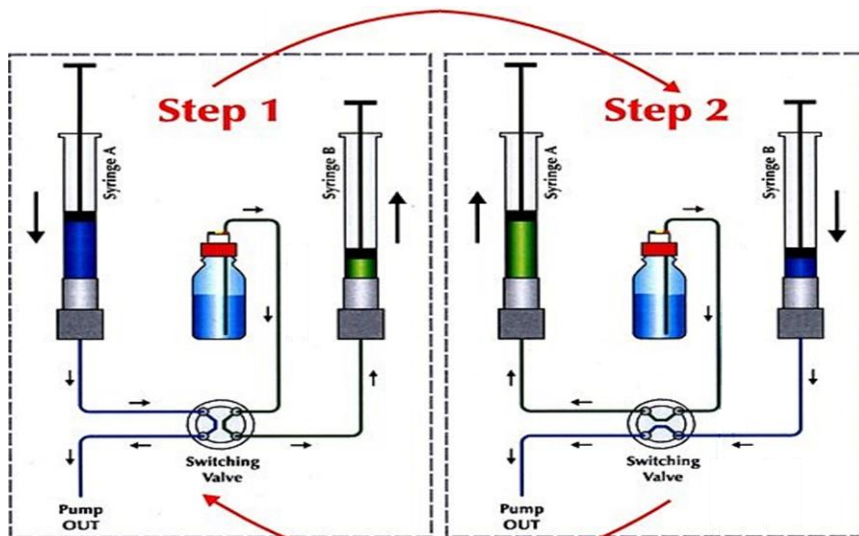
Several parameters are needed to be adjusted in order to miniaturize **HPLC** systems & to be suitable for the dimensions of capillary columns

Mobile phase flow rates

Low mobile phase flow rates ($\mu\text{L}/\text{min}$, nL/min) scale.



Pulsation-free
for flow rates
 $< 100 \mu\text{L}/\text{min}$



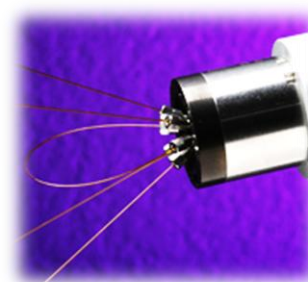
Syringe pumps

Sample injection volume

Injection volume accuracy and reproducibility.



Analytical injector
 $10\text{--}100 \mu\text{L}$
sample loop



Nanovolume injector
& switching valve

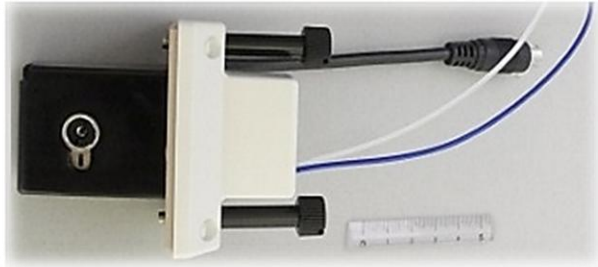
$4\text{--}20$
nanoliters



Internal sample
injectors

External injectors (VICI external injector – manual or automatic injector).

Detector cell volume



Standard analytical flow cell
5–10 mm path length,
2.5–18 μL cell volume

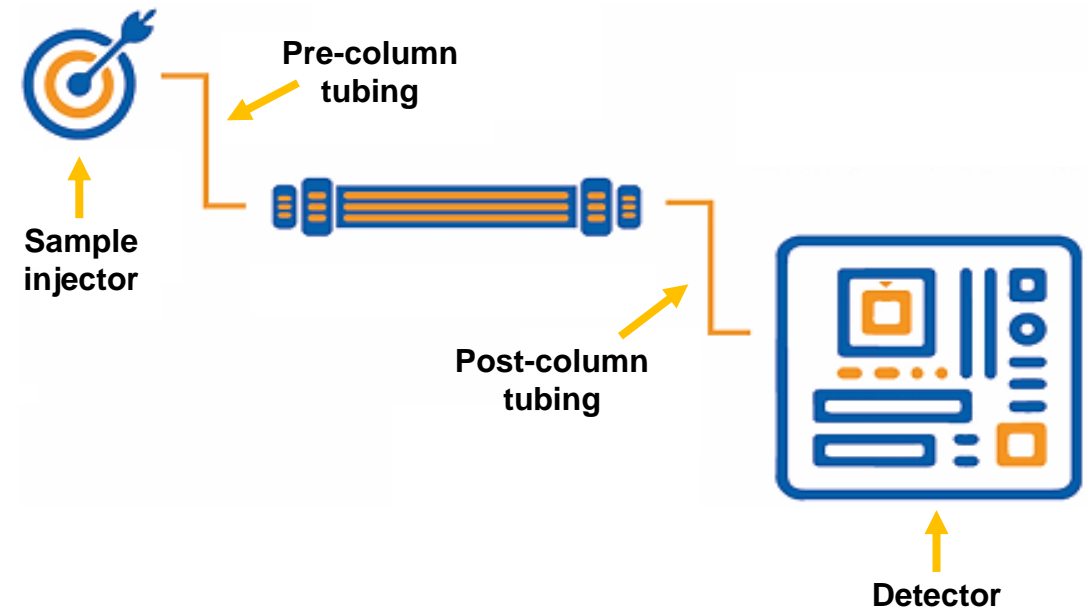
Nano flow cell
3–20 mm path length,
< 20 nL cell volume



Detector cell volume in the **UV/Vis** or spectroscopic photo detectors.

Extra column volume

Pre-column and post-column tubings and connections are very influential in these scales.



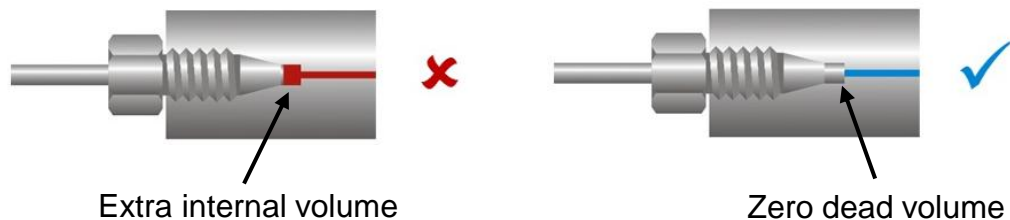
Extra-column volume must be minimized.

Fittings & connections

Fittings and connections at the downscale **LC** are not straightforward. The impact of column connection on band broadening.



How to achieve a perfect connection?



Conventional analytical scale (outer diameter) almost standard in all companies (1/16 inch).

Solvents leakage



Leaks can occur anywhere within the **HPLC**

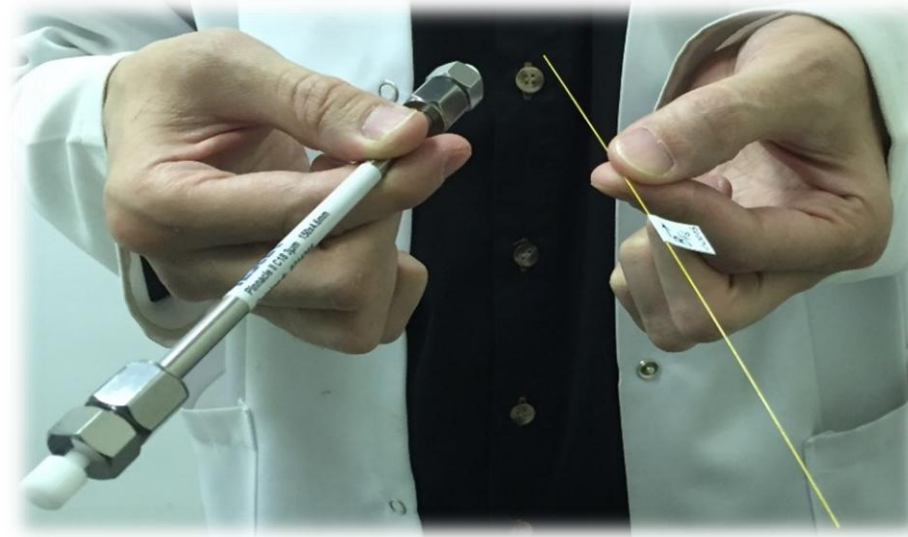
Leaks at the fittings	Leaks at the pump
Leaks at the injector	Leaks at the column
Leaks in the detector	

Difficult to detect mobile phases leakage.

Miniaturized LC system

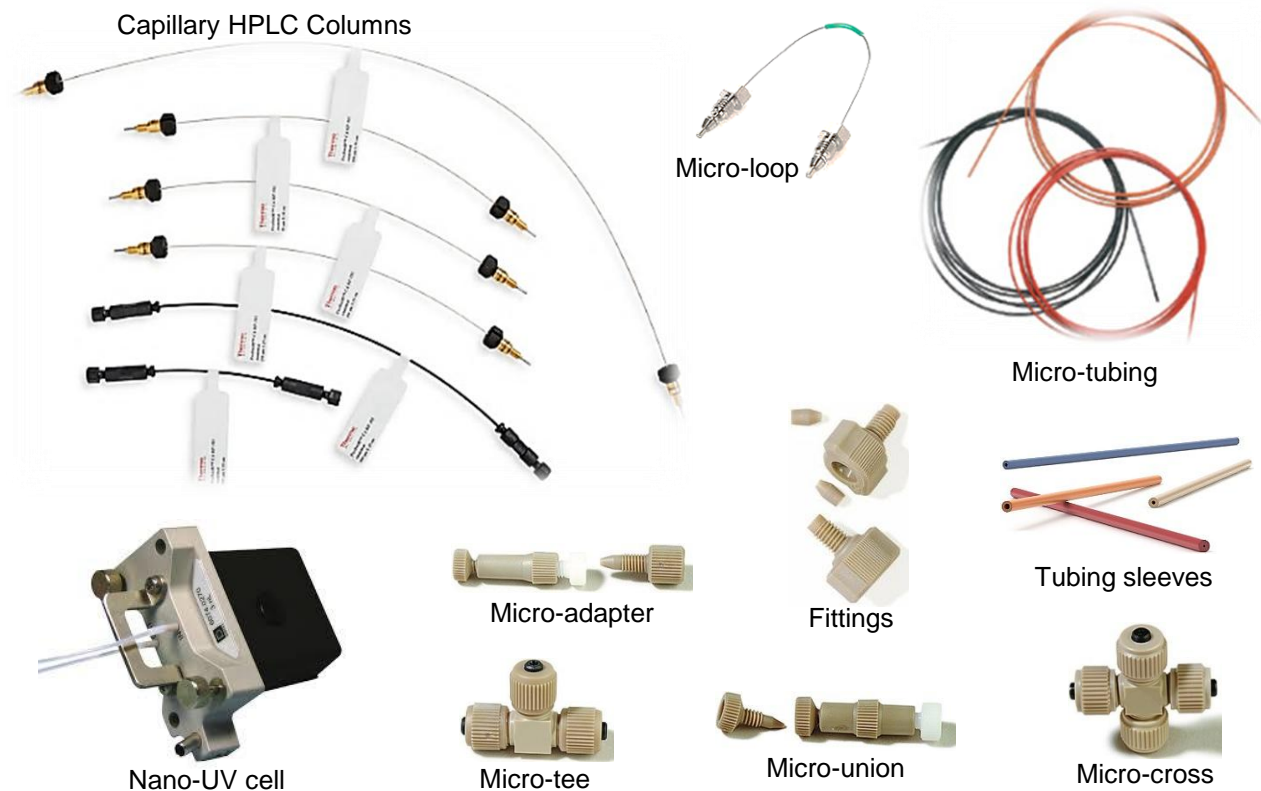
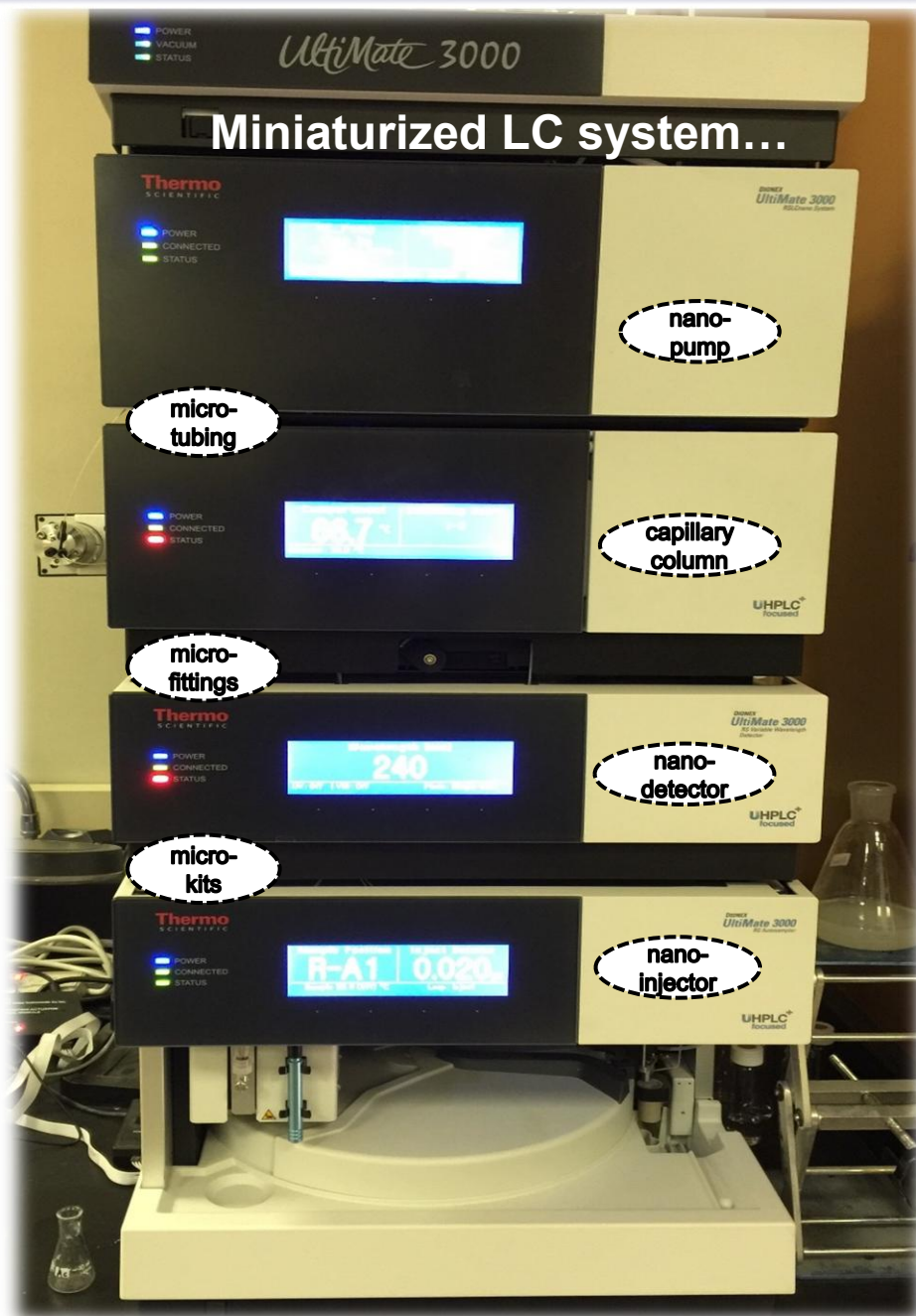


Commercial **nanoLC** systems



Conventional **LC** modification

-
- Mejía-Carmona K., et al., Trends in Analytical Chemistry 122 (2020) 115735.
 - Rieux L., et al., LCGC North America 29 (2011) 926.
 - Swart R., LC packings / Dionex, Netherlands, Chromedia 2008.



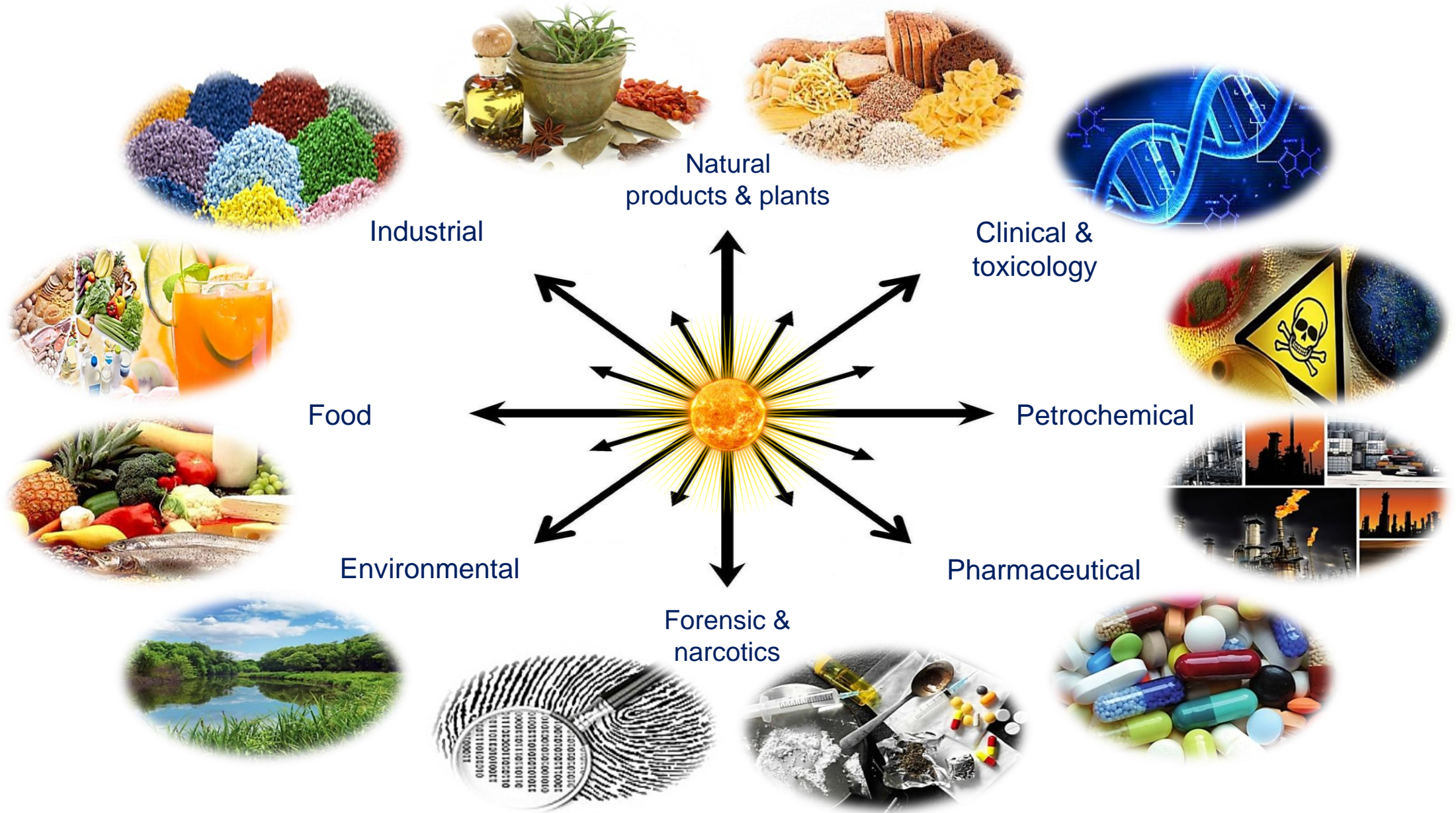
Miniaturized liquid chromatographs commercially available

Manufacturer	Brand name	Flow ($\mu\text{L}/\text{min}$)	Pressure (bar)
Waters	Acquity UPLC M-Class	0.2–100	1034
	nanoAcquity UPLC	0.2–100	690
Agilent	1260 Infinity nanoflow LC	0.1–1.0	400
	1200 Series capillary	1–100	400
Shimadzu	Prominence LC-20AD nano	0.001–5.0	400
Thermo	EASY-nano LC 1200	0.002–2.0	1200
	Dionex™ UltiMate 3000 series	0.001–50	800
AB Sciex	Ekspert™ nanoLC 400	0.1–50	690



Instrument
Columns
Accessories
Maintenance

Miniaturized LC Applications





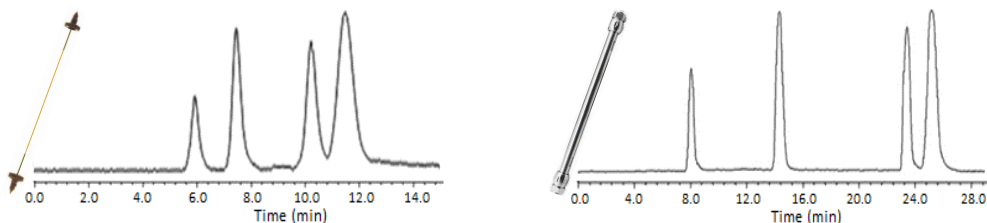
C₁₈ conventional column

Vs.



Capillary monolithic column

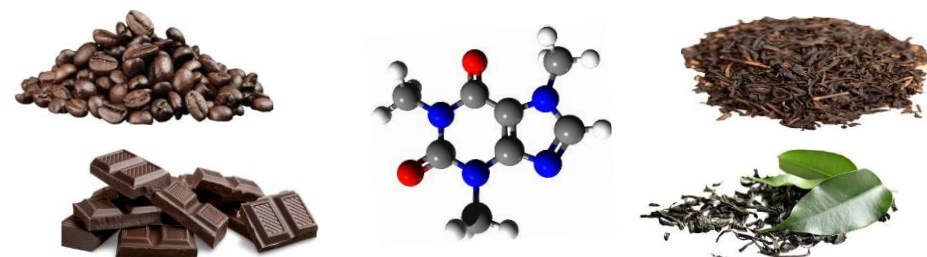
Determination of **BTEX** in water samples



	C₁₈ commercial column	Monolith capillary column
Column dimension	100 x 4.0 mm, 3 μ m	200 x 0.10 mm
Flow rate	500 μ L/min	0.40 μ L/min
Injection volume	10 μ L	4.0 nL
Detector cell volume	10 μ L	3.0 nL
Analysis time	27 min	12 min
N/m (tol)	48,900	33,400
R_s (o-xyl – m/p-xyl)	1.78	1.33
LOD (tol)	0.05 μ g/mL	0.01 μ g/mL
LOL (tol)	200 μ g/mL	150 μ g/mL
Sampling rate	\approx 2 sample/h	\approx 5 sample/h
Solvents consumption	30.0 mL/h	0.024 mL/h

-Aqel et al, Curr Anal Chem 16 (2020) 223

Determination of **caffeine** in food samples



	C₁₈ commercial column	Monolith capillary column
Column dimension	150 x 4.6 mm, 3 μ m	150 x 0.53 mm
Flow rate	500 μ L/min	41 μ L/min
Retention time	5.78 min	1.16 min
Detector cell volume	10 μ L	1.0 μ L
Injection volume	10 μ L	1.0 μ L
Plates number	28,400	12,700
LOD	0.20 μ g/mL	0.05 μ g/mL
LOQ	0.70 μ g/mL	0.16 μ g/mL
LOL	150 μ g/mL	250 μ g/mL
Sampling rate	\approx 10 h ⁻¹	\approx 40 h ⁻¹
Solvents consumption	30.0 mL/h	2.46 mL/h

-ALothman et al, Food Chem 132 (2012) 2217

Thank You!

