

المعمل الثامن:

تصميم الباديء لتفاعل إنزيم البلمرة التسلسلي

Designing PCR Primers

251Mbio

Amal Alghamdi

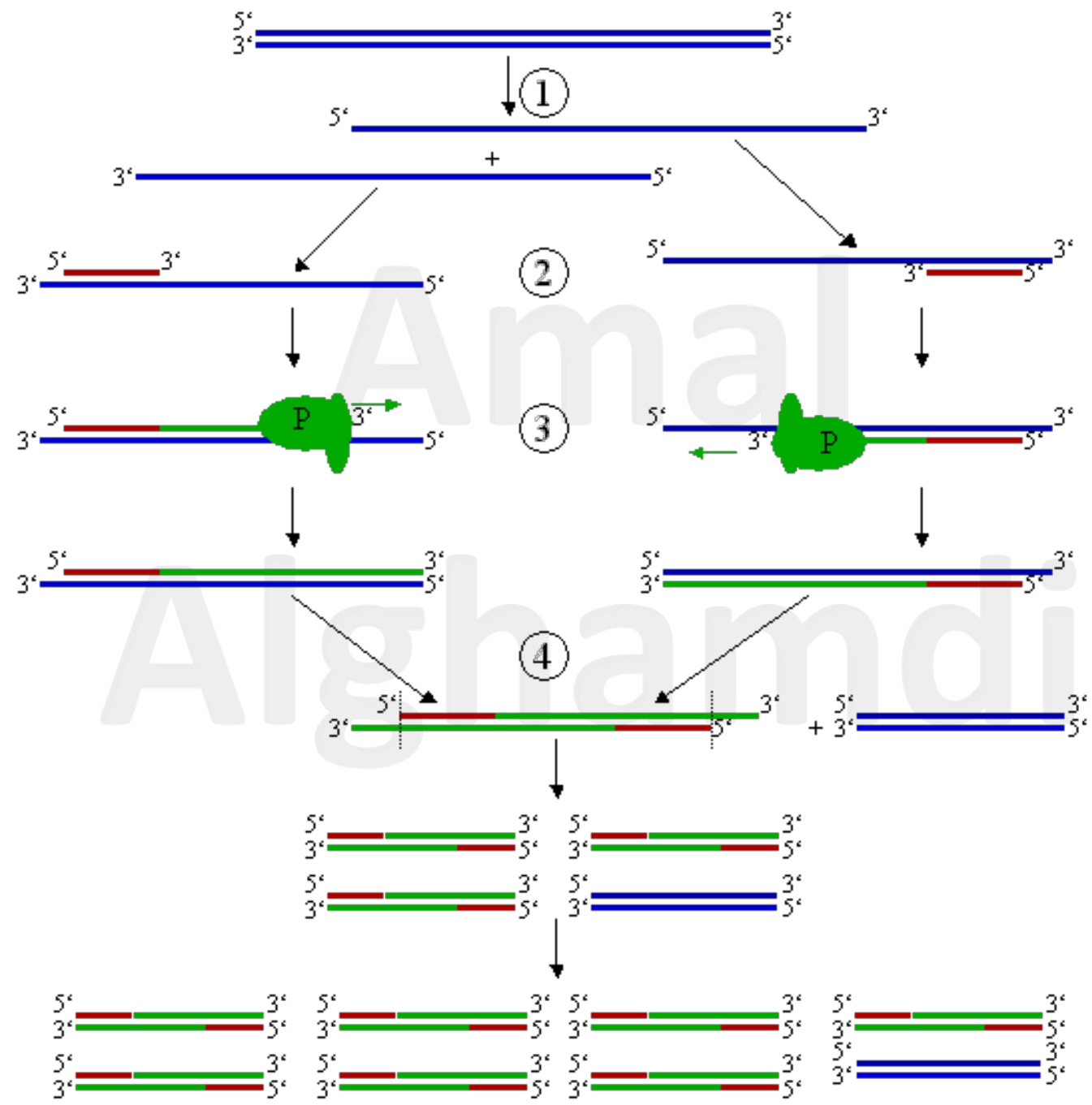
2018

Terminologies بعض المصطلحات الهامة

- الباديء Primer .
- تتابع القالب Template sequence .
- درجة الذوبان (Tm) Melting Temperature .
- درجة الارتباط (Ta) Annealing Temperature .
- تفاعل انزيم البلمرة التسلسلي PCR .

الأنواع المختلفة لتفاعل إنزيم البلمرة التسلسلي

- Standard PCR.
- Nested PCR
- Touch down PCR
- Sequencing PCR
- Intersequence--specific PCR (ISSR)
- والكثير غيرها



مصدر التتابع القالب

Origin of the Template Sequence

- تتابع خاص بالباحث Self-generated.
- تتابع نيوكليوتيدي من قاعدة بيانات التتابعات النيوكليوتيدية Nucleotide Sequence Databases مثل (NCBI's و GenBank).
- الحصول على التتابع الصحيح من قاعدة البيانات
- ينسخ التتابع في قاعدة البيانات لإستخراج التتابعات المشابهة من نفس الكائن الحي تحت الدراسة.

ما هو الباديء Primer؟

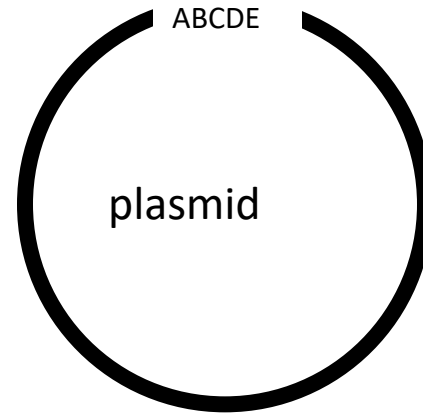
- هو تتابع قصير من النيوكليوتيدات، يتم تصميمه طبقاً لتتابع معاكس ومكمل لمنطقة محددة من جينوم الحمض النووي DNA المستهدفة.

Alghamdi

prepare PCR product
prepare vector



PCR product

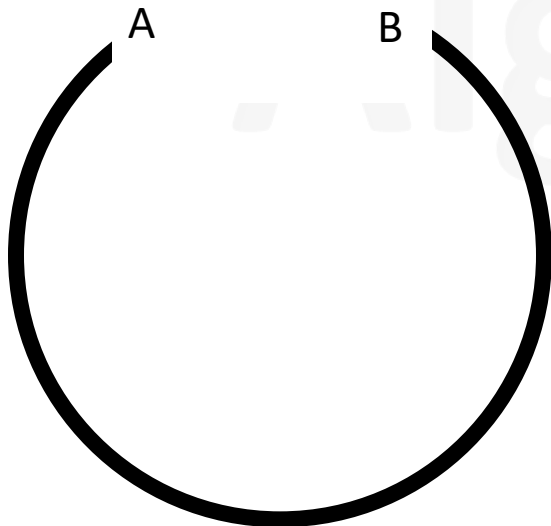


plasmid



B

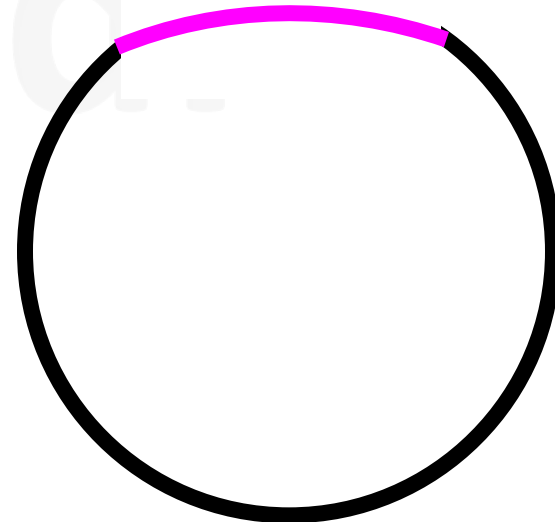
restriction
digest



A

B

ligate



أهم مواصفات تتابع الباديء Primer

- يتراوح طوله بين 18 – 28 نيوكليوتيدة.
- نسبة GC له 50-60 %.
- يحتوي توزيع متوازن من القواعد النيتروجينية G/C و A/T.

تتابع صالح

5' ATGCACTCAGACGTACAACG
TGAC 3'
24 bases
AT: 12
GC: 12 (50% GC)
Balanced distributon
Ta = 65°C

تتابع غير صالح

5' AACAAACGATTTTTT 3'
17 bases
AT: 14
GC: 3 (18 % GC)
Unbalanced distributon
Ta= 38°C

الغرض من تصميم الباديء Purpose

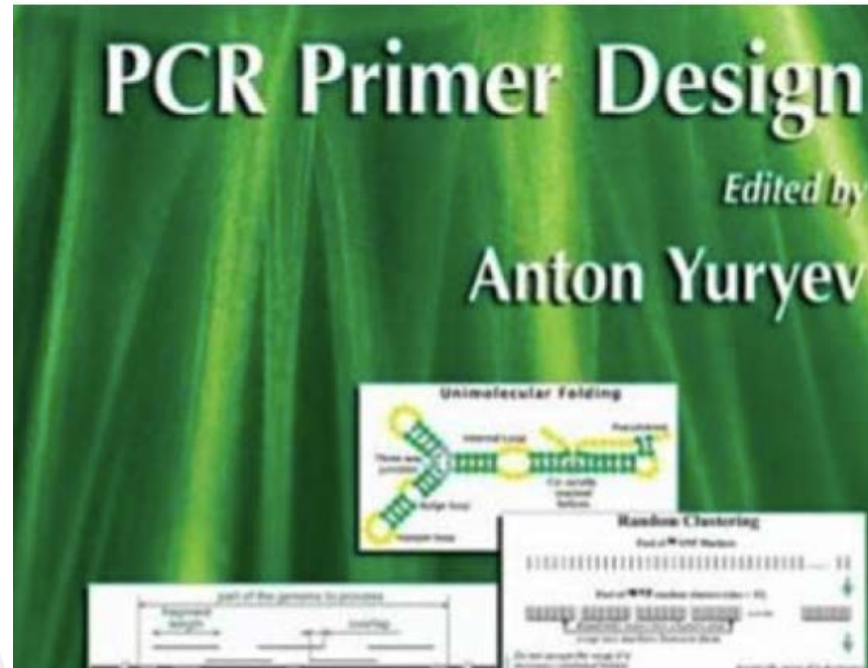
- لغرض الكلونة Gene cloning.
- للإختبارات التشخيصية باستخدام تقنية PCR.
- للتقدير الكمي للجينات أو نسخ الحمض النووي RNA (Transcript).

مصادر تصميم الباديء على الإنترنت

Sources of Primer Design

- CLC Workbench
- Primer3
- Primer3 Plus
- PrimerZ
- PerlPrimer
- (Google) وغيره الكثير بالبحث في محرك البحث

المرجع Reference



- **Freely accessible Primer Design Book**
 - Explains the design of complex primers blow by blow:

http://vetbiotech.um.ac.ir/parameters/vetbiotech/filemanager/new_admin/books/PCR%20Primer%20Design.pdf

مثال: جين lacZ gene في جينوم بكتيريا Escherichia coli

- يحتوي البلازميد pUC18 على تتابع لجين Lac Z الذي يشفر إنزيم الجالاکتوسيداز b- galactosidase.
- يحتوي هذا البلازميد على جين المقاومة للمضاد الحيوي Ampicillin.

مثال: جين lacZ gene في جينوم بكتيريا Escherichia coli

5' end of lacZ gene

3' end of lacZ gene

5' atgaccatga ttacggattc actggccgtc
3' tactggtact aatgcctaag tgaccggcag

5' gctaccatta ccagttggtc tgggtgtcaaa aataa 3'
3' cgatggtaat ggtcaaccag accacagttt ttatt 5'

atg acc atg att acg gat tca ctg gcc gtc
M T M I T D S L A V

cag ttg gtc tgg tgt caa aaa taa
Q L V W C Q K *

lacZ beta-D-galactosidase [*Escherichia coli* str. K-12 substr. MG1655]

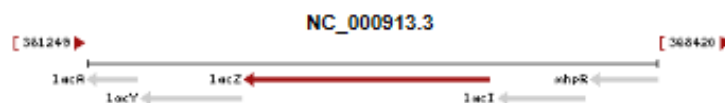
Gene ID: 945006, updated on 4-Feb-2018

Summary

Gene symbol	lacZ
Gene description	beta-D-galactosidase
Primary source	EcoGene:EG10527
Locus tag	b0344
Gene type	protein coding
RefSeq status	PROVISIONAL
Organism	Escherichia coli str. K-12 substr. MG1655 (strain: K-12, substrain: MG1655)
Lineage	Bacteria; Proteobacteria; Gammaproteobacteria; Enterobacterales; Enterobacteriaceae; Escherichia
Also known as	ECK0341; JW0335
Summary	Repressed during biofilm formation. [More information is available at EcoGene: EG10527]. galactosidase is a metalloenzyme exhibiting broad substrate specificity. [More information is available at EcoCyc: EG10527].

Genomic context

Sequence: NC_000913.3 (383231..386305, complement)



Genomic regions, transcripts, and products

Genomic Sequence: NC_000913.3

Go to [reference sequence details](#)Go to nucleotide: [Graphics](#) [FASTA](#) [GenBank](#)

1- البحث عن التتابع النيوكليوتيدي للجين PuC 18 :

- تم اختيار **Synthetic construct DNA, plasmid vector pUC18 including** (GenBank: LC129268.1) **artificial sequence**.
- <https://www.ncbi.nlm.nih.gov/nucore/LC129268.1?&feature=any>

- بعد اختيار FASTA يتم اختيار highlight sequence feature
- يتم تظليل exons الخاصة بالجين
- يتم نسخ التتابعات المظلة ثم نقلها إلى Primer3web

Synthetic construct DNA, plasmid vector pUC18 including artificial sequence

GenBank: LC129268.1

[FASTA](#) [Graphics](#)

[Go to:](#) ☑

LOCUS LC129268 2808 bp DNA linear SYN 19-MAR-2016
DEFINITION Synthetic construct DNA, plasmid vector pUC18 including artificial sequence.
ACCESSION LC129268
VERSION LC129268.1
KEYWORDS .
SOURCE synthetic construct
ORGANISM [synthetic construct](#)
other sequences; artificial sequences.
REFERENCE 1
AUTHORS Takahashi,M., Kita,Y., Mizuno,A. and Goto-Yamamoto,N.
TITLE Evaluation of method bias in bacterial community analysis
JOURNAL Unpublished
REFERENCE 2 (bases 1 to 2808)
AUTHORS Takahashi,M. and Goto-Yamamoto,N.
TITLE Direct Submission
JOURNAL Submitted (03-MAR-2016) Contact:Masayuki Takahashi National Research Institute of Brewing, Technology Development Research Division; 3-7-1 Kagamiyama, Higashi-hiroshima, Hiroshima 739-0046, Japan URL :<http://www.nrrib.go.jp/index.html>

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121 ttggcgggtg tcggggctgg cttactatg cggcatcaga cgagattgta ctgagagtg
181 accatatgcg gtgtgaaata ccgcacagat cgtaaggag aaaaaccgc atcaggcgc
241 attgccatt caggctcgc aactgttggg aaggcgcgc ggtcgggcc tcttgcctat
301 tacgccagct ggcgaaaggg ggatgtgct caaggcgatt aagtgggta acgccagggt
361 ttcccaatc acgacattgt aaaaacacgc ccaatgccaa gcttgcctgc ctacgatgc

Customize view ▾

Analyze this sequence ▴

[Run BLAST](#)

[Pick Primers](#)

[Highlight Sequence Features](#)


[Find in this Sequence](#)


Related information ▴


[Taxonomy](#)

Recent activity ▴

[Turn Off](#) [Clear](#)

 Synthetic construct DNA, plasmid vector pUC18 including artificial sequence Nucleotide

 pUC18 vector (1388) Nucleotide

 pUC18 cloning vector Nucleotide

 Highlight Sequence Features - NCBI Video Vault

 pUC18 (10816) Nucleotide

[See more...](#)

439..560
/note="internal standard DNA for quantification of microbial rDNA using quantitative PCR"

NCBI Resources ☑ How To ☑

Nucleotide Nucleotide ▾ [Help](#)

FASTA ▾

Synthetic construct DNA, plasmid vector pUC18 including artificial sequence

GenBank: LC129268.1

[GenBank](#) [Graphics](#)

>LC129268.1:439-560 Synthetic construct DNA, plasmid vector pUC18 including artificial sequence

AACTAATACGACTCACTATAGGGTCCGATCTCCGAGGCTCATATCGATCGGTAGGGCATCTAATGGCTTCGGAGTCAAGGGCTATATTCGCCATGTGATGTCGAAAGCCGG

Send to: ▾

Change region shown ▴

Whole sequence

Selected region

from: 439 to: 560

Customize view ▾

Analyze this sequence ▴

[Run BLAST](#)

[Pick Primers](#)

[Highlight Sequence Features](#)

[Find in this Sequence](#)

Related information ▴

[Taxonomy](#)

Nucleotide

GenBank

E. coli gene lacZ coding for beta-galactosidase (EC 3.2.1.23)

GenBank: V00296.1

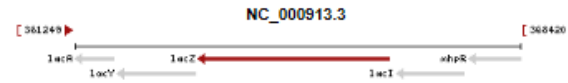
[FASTA](#) [Graphics](#)

Go to:

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 VERSION V00296.1
 KEYWORDS galactosidase.
 SOURCE Escherichia coli
 ORGANISM [Escherichia coli](#)
 Bacteria; Proteobacteria; Gammaproteobacteria; Enterobacterales; Enterobacteriaceae; Escherichia.
 REFERENCE 1 (bases 1 to 3078)
 AUTHORS Kalnins,A., Otto,K., Ruther,U. and Muller-Hill,B.
 TITLE Sequence of the lacZ gene of Escherichia coli
 JOURNAL EMBO J. 2 (4), 593-597 (1983)
 PUBMED [6313347](#)
 REFERENCE 2
 AUTHORS Zell,R. and Fritz,H.J.
 TITLE DNA mismatch-repair in Escherichia coli counteracting the hydrolytic deamination of 5-methyl-cytosine residues
 JOURNAL EMBO J. 6 (6), 1809-1815 (1987)
 PUBMED [3038536](#)
 COMMENT Data kindly reviewed (18-MAY-1983) by U. Ruether.
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Genomic context

Sequence: NC_000913.3 (363231..366305, complement)

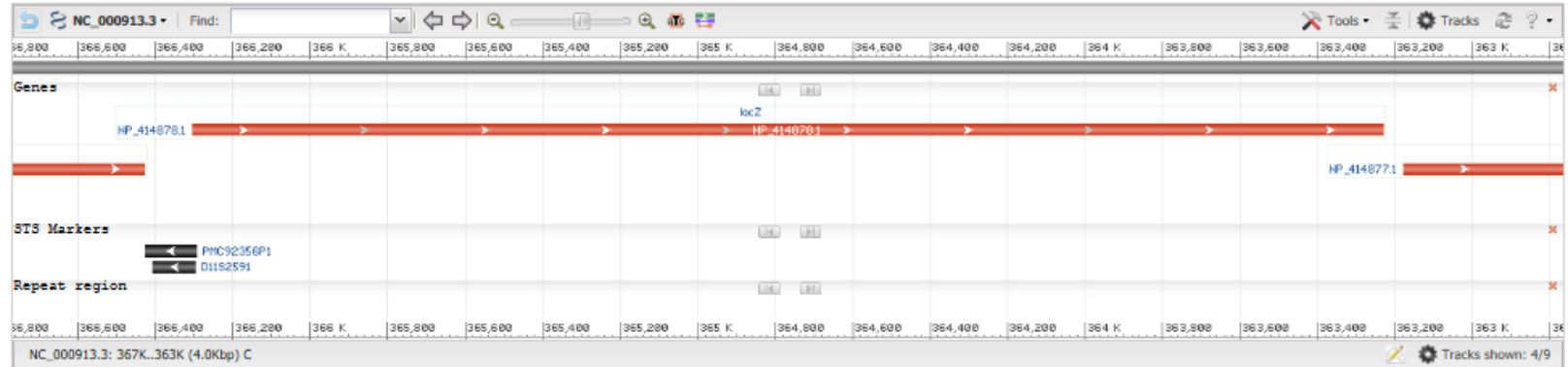


Genomic regions, transcripts, and products

Genomic Sequence: NC_000913.3

[Go to reference sequence details](#)

[Go to nucleotide: Graphics FASTA GenBank](#)



Bibliography

FASTA - Send to: -

Escherichia coli str. K-12 substr. MG1655, complete genome

NCBI Reference Sequence: NC_000913.3

[GenBank](#) [Graphics](#)

>NC_000913.3:c366305-363231 Escherichia coli str. K-12 substr. MG1655, complete genome
 ATGACCATGATTACGGATTCACTGGCCGTGGTTTACAAGTCGTGACTGGGAAACCCCTGGCGTTACCC
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Change region shown

Whole sequence

Selected region

from: 363231 to: 366305

Update View

Customize view

Display options

Show reverse complement

Update View

Analyze this sequence

Run BLAST

Pick Primers

Highlight Sequence Features

Find in this Sequence

Related information

Assembly

BioProject

BioSample

Protein

PubMed

Taxonomy

Components (Core)

Full text in PMC

ORIGIN

1 accatgatta cggattcact gcccctgctt ttacaacgct gtagctggga aaacctggc
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 2521 cgtgagcag atcagggaa aacctatt atcagccga aaacctacc gattgatgt

2- النسخ في Primer3web

- يتم لصق التتابع الخاص في المكان المخصص
- يتم اختيار Pick Primer
- تظهر تتابع الباديء الأمامي والعكسي.

• http://primer3.ut.ee/cgi-bin/primer3/primer3web_results.cgi

Primer3 Input

[primer3.ut.ee/](#)

All rights reserved. This file is part of the primer3web suite. The primer3 suite and libraries are free software; you can redistribute them and/or modify them under the terms of the GNU General Public License as published by the Free Software Foundation; either version 2 of the License, or (at your option) any later version.

[Department of Bioinformatics](#) · [Primer3Web Release 4.1.0 ...](#) · [Primer3 k-mer lists](#)

People also search for

[primer3 manual](#) [netprimer](#)
[primer3 download](#) [ucsc in silico pcr](#)
[snp check](#) [primerblast](#)

Primer3 Input (version 0.4.0)

[bioinfo.ut.ee/primer3-0.4.0/](#)

There is a newer version of Primer3 available at <http://primer3.ut.ee>. Paste source sequence below (5'->3', string of ACGTNaagtn -- other letters treated as N ...

Run Primer3Web - Primer3Plus

https://primer3plus.com/primer3web/primer3web_input.htm

Pick left primer, or use left primer below. Pick hybridization probe (internal oligo), or use oligo below. Pick right primer, or use right primer below (5' to 3' on opposite strand). Sequence Id, A string to identify your output. Targets, E.g. 50,2 requires primers to surround the 2 bases at positions 50 and 51. Or mark the source ...

Primer3Plus

<https://primer3plus.com/>

Primer3 Development. On this place the efforts of developing Primer3, Primer3Plus and Primer3Web are coordinated. Please go there for bug reports, feature requests and support.

Amal

Primer3web version 4.1.0 - Pick primers from a DNA sequence.

[disclaimer](#)

[code](#)

[cautions](#)

Select the [Task](#) for primer selection

[Template masking before primer design \(available species\)](#)

Select species

Nucleotides to mask in 5' direction

Primer failure rate cutoff

Nucleotides to mask in 3' direction

Paste source sequence below (5'->3', string of ACGTNaagtn -- other letters treated as N -- numbers and blanks ignored). FASTA format ok. Please N-out undesirable sequence (vector, ALUs, LINEs, etc.) or use a [Mispriming Library \(repeat library\)](#)

Pick left primer, or use left primer below Pick hybridization probe (internal oligo), or use oligo below Pick right primer, or use right primer below (5' to 3' on opposite strand)

[Sequence Id](#)

A string to identify your output.

[Targets](#)

E.g. 50,2 requires primers to surround the 2 bases at positions 50 and 51. Or mark the [source sequence](#) with [and]: e.g. ...ATCT[CCCC]TCAT.. means that primers must flank the central CCCC.

[Overlap Junction List](#)

E.g. 27 requires one primer to overlap the junction between positions 27 and 28. Or mark the [source sequence](#) with -: e.g. ...ATCTAC-TGTCAT.. means that primers must overlap the junction between the C and T.

[Excluded Regions](#)

E.g. 401,7 68,3 forbids selection of primers in the 7 bases starting at 401 and the 3 bases at 68. Or mark the [source sequence](#) with < and >: e.g. ...ATCT<CCCC>TCAT.. forbids primers in the central CCCC.

[Pair OK Region List](#)

See manual for help.

[Included Region](#)

E.g. 20,400: only pick primers in the 400 base region starting at position 20. Or use { and } in the [source sequence](#) to mark the beginning and end of the included region: e.g. in ATC{TTC...TCT}AT the included region is TTC...TCT.

Select the [Task](#) for primer selection

Template masking before primer design (available species)	
Select species <input type="text" value="Example: Mus musculus"/>	Nucleotides to mask in 5' direction <input type="text" value="1"/>
Primer failure rate cutoff < <input type="text" value="0.1"/>	Nucleotides to mask in 3' direction <input type="text" value="0"/>

Paste source sequence below (5'→3', string of ACGTNacgtn -- other letters treated as N -- numbers and blanks ignored). FASTA format ok. Please N-out undesirable sequence (vector, ALUs, LINEs, etc.) or use a [Mispriming Library \(repeat library\)](#)

```
1 accatgatta cggattcact ggccgtcgtt ttacaacgtc gtgactggga aaaccctggc
  61 gttaccacaac ttaatgcct tgcagcacat ccccctttcg ccagctggcg taatagcga
 121 gaggcccgca cggatcgccc ttcccaacag ttgcgcagc tgaatggcga atggccttt
 181 gcctggtttc cggcaccaga agcgggtccg gaaagctgac tggagtgcga tcttcctgag
 241 gccgatactg tcgtcgtccc ctcaaacagg cagatgcac gttacgatgc gcccatctac
 301 accaacgtaa cctatccat tacgtcaat ccgctgttg ttcccaacga gaatccgacg
```

<input checked="" type="checkbox"/> Pick left primer, or use left primer below	<input type="checkbox"/> Pick hybridization probe (internal oligo), or use oligo below	<input checked="" type="checkbox"/> Pick right primer, or use right primer below (5' to 3' on opposite strand)
<input type="text"/>	<input type="text"/>	<input type="text"/>

- [Sequence Id](#) A string to identify your output.
- [Targets](#) E.g. 50,2 requires primers to surround the 2 bases at positions 50 and 51. Or mark the [source sequence](#) with [and]: e.g. ...ATCT[CCCC]TCAT.. means that primers must flank the central CCCC.
- [Overlap Junction List](#) E.g. 27 requires one primer to overlap the junction between positions 27 and 28. Or mark the [source sequence](#) with -: e.g. ...ATCTAC-TGTCAT.. means that primers must overlap the junction between the C and T.
- [Excluded Regions](#) E.g. 401,7 68,3 forbids selection of primers in the 7 bases starting at 401 and the 3 bases at 68. Or mark the [source sequence](#) with < and >: e.g. ...ATCT<CCCC>TCAT.. forbids primers in the central CCCC.
- [Pair OK Region List](#) See manual for help.
- [Included Region](#) E.g. 20,400: only pick primers in the 400 base region starting at position 20. Or use { and } in the [source sequence](#) to mark the beginning and end of the included region: e.g. in ATC{TTC...TCT}AT the included region is TTC...TCT.
- [Start Codon Position](#)
- [Internal Oligo Excluded Region](#)
- [Force Left Primer Start](#) [Force Right Primer Start](#)
- [Force Left Primer End](#) [Force Right Primer End](#)

[Sequence Quality](#)

Primer3 Output

WARNING: Numbers in input sequence were deleted.

PRIMER PICKING RESULTS FOR

Template masking not selected

No mispriming library specified

Using 1-based sequence positions

OLIGO	start	len	tm	gc%	any_th	3'_th	hairpin	seq
LEFT PRIMER	26	20	58.04	55.00	0.97	6.85	0.00	acgactcactatagggtccg
RIGHT PRIMER	130	21	58.01	47.62	0.00	0.00	0.00	gcatacaaatctgacatggcg

SEQUENCE SIZE: 150

INCLUDED REGION SIZE: 150

PRODUCT SIZE: 105, PAIR ANY_TH COMPL: 0.00, PAIR 3'_TH COMPL: 0.00

```
1 actctagaggatccccggaactaatacagactcactatagggtccgatcttccgaggtctc
  >>>>>>>>>>>>>>>>>>>>>
```

```
61 atatcgatcggtagggcatctaattggcttcggagttcaagggttatattcgccatgtcag
  <<<<<<<<<<<<<<
```

```
121 atttgatatgccaaaggccgggtaccgagct
  <<<<<<<<<<
```

KEYS (in order of precedence):

>>>>> left primer

<<<<<< right primer

