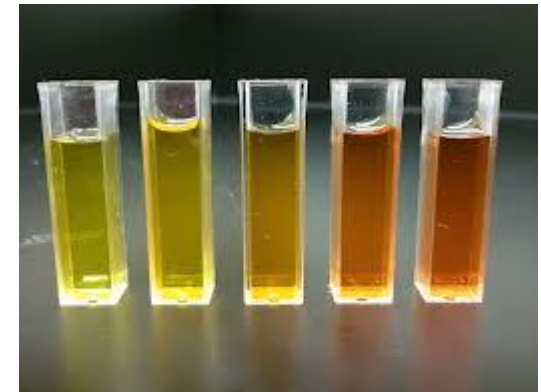


Kinetics analysis of
 β -fructofuranosidase enzyme

4-The effect of sucrose concentration on the rate of reaction catalyzed by β -fructofuranosidase enzyme .



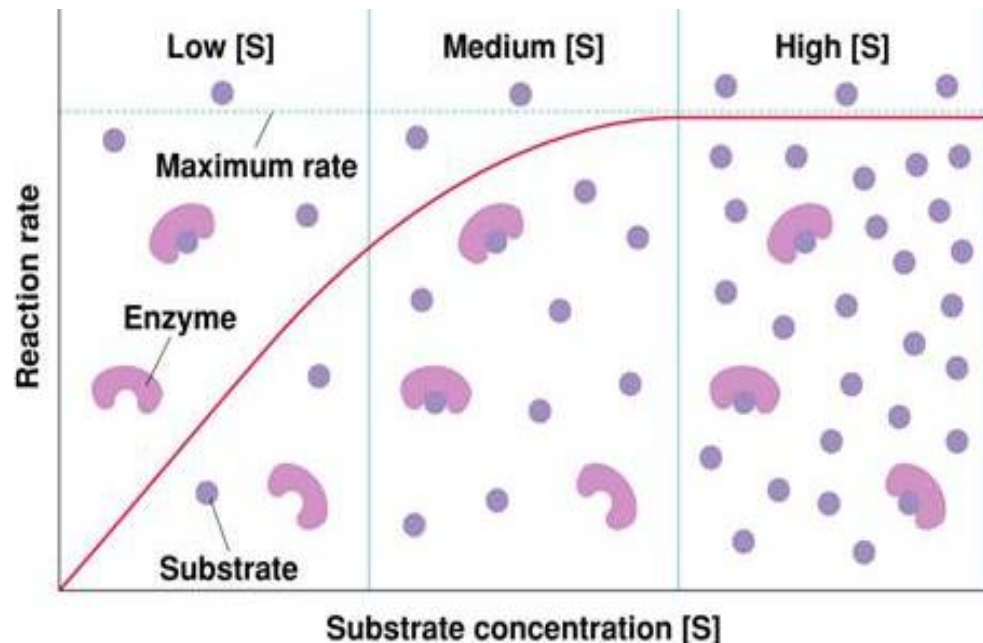
- One of the important parameter affecting the rate of a reaction catalyzed by an enzyme is the substrate concentration, [S].
- During enzyme substrate reaction, the initial velocity V_0 gradually increases with increasing the concentration of the substrate. Finally a point is reached, beyond which the increase in V_0 will not depend on the [S].
- When a graph plotted with substrate concentration on the X axis and corresponding velocity on Y axis. It can be observed from the graph that as the concentration of the substrate increases, there is a corresponding increase in the V_0 .
- However beyond a particular substrate concentration, the velocity remains constant without any further increase. This maximum velocity of an enzyme catalyzed reaction under substrate saturation is called the V_{\max} , Maximum velocity.

As the substrate level is increased , the velocity increases in a hyperbolic fashion.

At relatively low concentration of substrate, the rate of reaction increase linearly with an increase in substrate concentration.

At higher substrate concentration the rate of reaction increase *smaller and smaller* amount in response to increase in substrate concentration.

Finally , a point is reached beyond which there is only small increase in the rate of the reaction with increasing substrate concentration. **This plateau is called maximum velocity, V_{max} .**



Michaelis –Menten Plot

The hyperbolic shape of this curve can be expressed algebraically by the Michaelis –Menten equation:

$$V_o = V_{\max} [S] / K_M + [S]$$

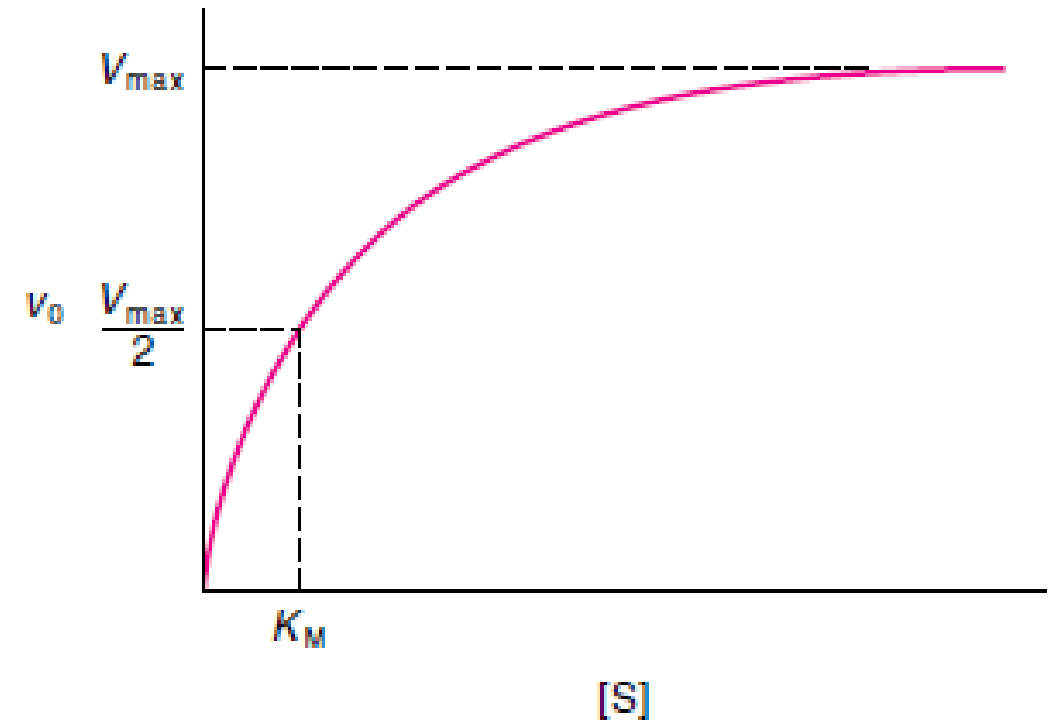
V_i = initial velocity

V_{\max} = maximum velocity

$[S]$ = substrate concentration

K_M = Michaelis -Menten constant

K_M : is a substrate concentration at half V_{\max} (K_M indicate the affinity of an enzyme for its substrate)



Note that the larger the K_M (the weaker the binding), the larger the $[S]$ needed to reach the half maximum rate.

The K_m can vary greatly from enzyme to enzyme , and even for different substrates of the same enzyme.

The Michaelis -Menten equation can be algebraically transformed into forms that are useful in the practical determination of K_m and V_{max} . One common transformation is derived simply **by taking the reciprocal of both sides of the Michaelis -Menten equation to give Lineweaver – Burk equation:**

$$\frac{1}{V_i} = \frac{1}{V_{max}} + \frac{K_m}{V_{max}} \cdot \frac{1}{[S]}$$

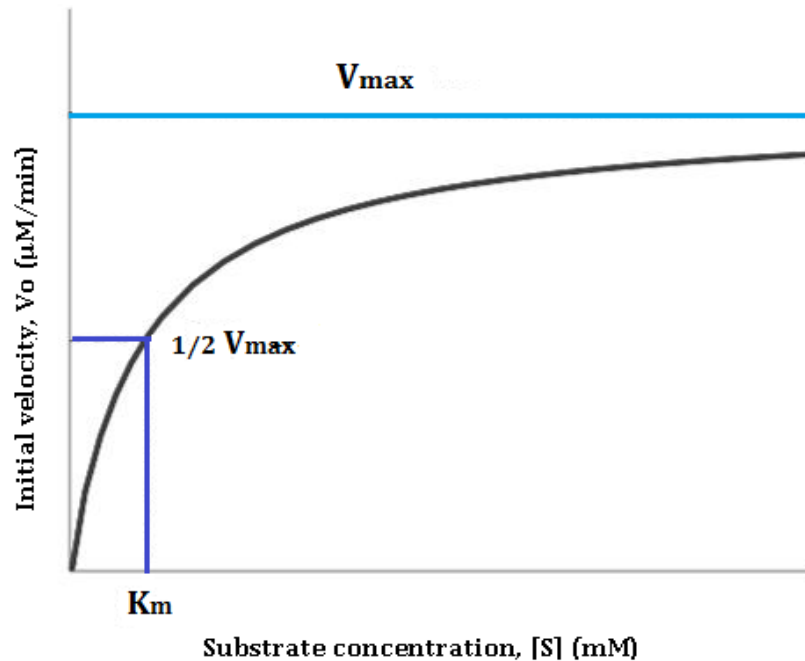
Lineweaver – Burk equation

Lineweaver – Burke plot

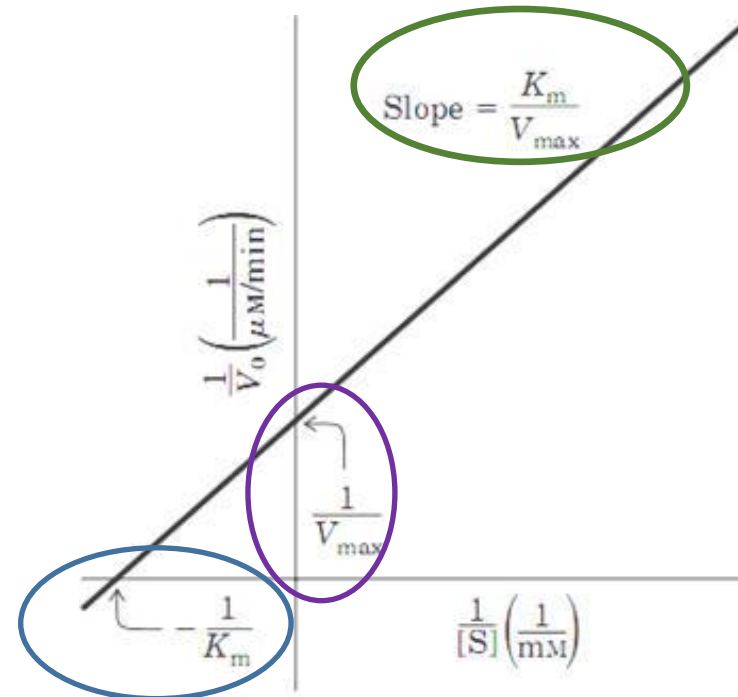
For enzymes obeying the Michaelis-Menten relationship, the “double reciprocal” of the V_0 versus $[S]$ from the first graph, (figure 1) yields a straight line (figure 2).

The **slope** of this straight line is K_M / V_{max} , which has an intercept of $1/V_{max}$ on the $1/V_0$ (y-axis), and an intercept of $-1/K_M$ on the $1/[S]$ (x-axis).

The main advantage of Lineweaver-Burk plot is to determine the V_{max} and K_M more accurately, which can only be approximated from **Michaelis – Menten** graph of V_0 versus $[S]$ (Fig 1).



(figure 1)



(figure 2)

Objectives:

- 1) To establish the relationship between substrate concentration and the rate of an enzyme catalyzed reaction.
- 2) To determine the K_m and V_{max} of the enzyme for a particular substrate.

Principle:

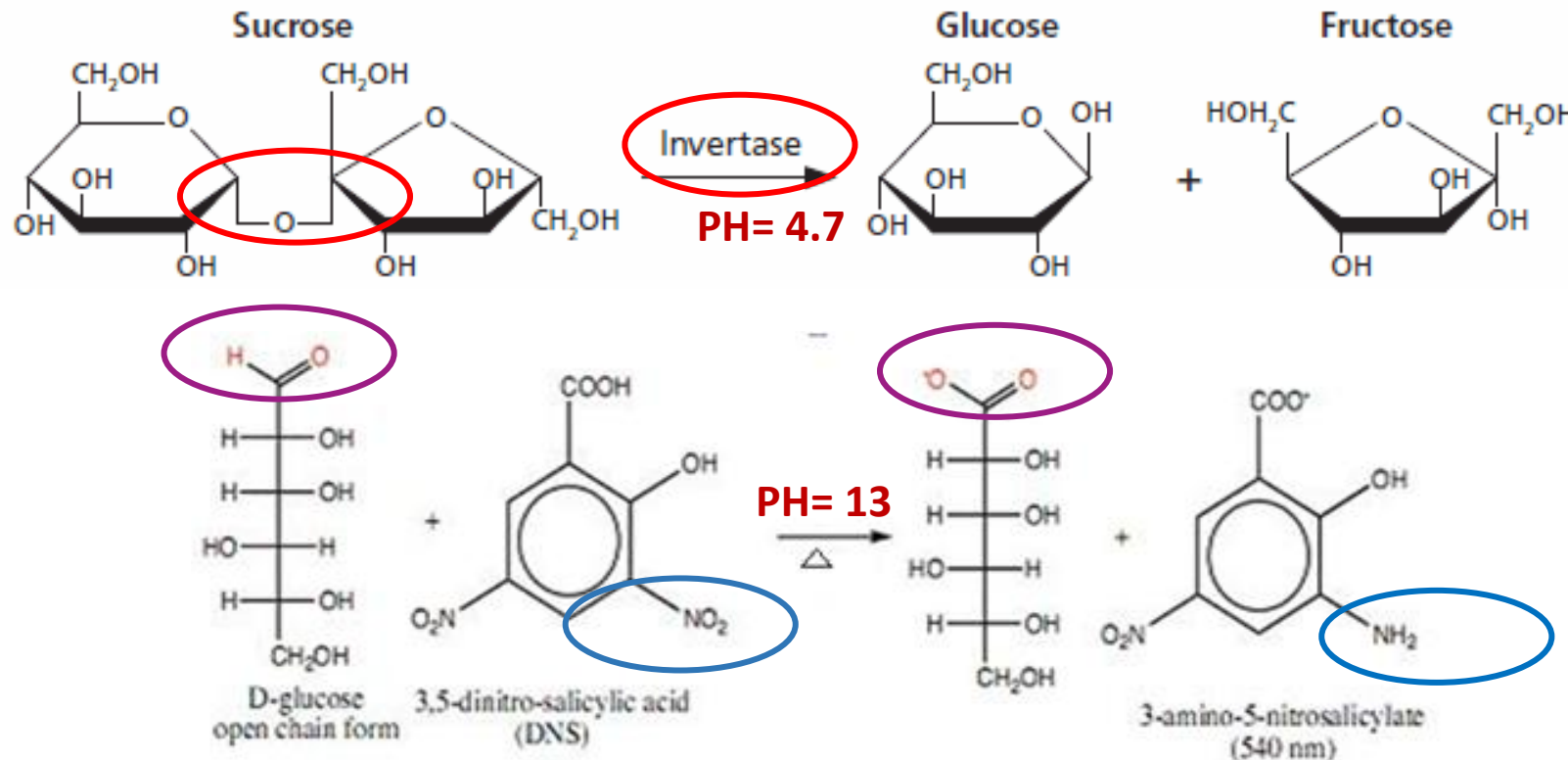
The enzyme **β -fructofuranosidase** can catalyze the hydrolysis of sucrose with the production of reducing sugars.

In the study of substrate concentration on enzyme kinetics, the enzyme is kept constant whereas the concentration of sucrose is taken in increasing order. As the substrate concentration increases, the amount of products produced in every tube also increases.

This was explained by Michaelis and others that an enzyme catalyzed reaction at varying substrate concentrations is **diphasic** i.e. at low substrate concentration the active sites on (enzyme) are not occupied by substrate and the enzyme rate varies with substrate molecules concentration (**phase I**).

As the number of substrate molecules increases, the enzyme reaches the saturation level, since there is no more reaction sites remaining for binding. So the enzyme can work with full capacity and its reaction rate is independent of substrate concentration. (**Phase II**).

This Enzyme – substrate reaction can be determined by measuring the increase in reducing sugars using the 3, 5 Dinitro salicylic acid reagent. In an alkaline condition, the pale yellow colored the 3, 5-dinitro salicylic acid undergo reduction to yield orange colored 3-amino -5-nitrosalicylic acid. The absorbance of resultant solutions is read at 540nm. The intensity of color depends on the concentration of reducing sugars produced.



Sucrose is hydrolyzed by invertase in an acidic media, then the reaction is inhibited by DNS, which has a high PH

DNS is an oxidizing agent and undergo reduction to form an amino group

Glucose and fructose is an reducing agent and undergo oxidation to form a carboxyl group

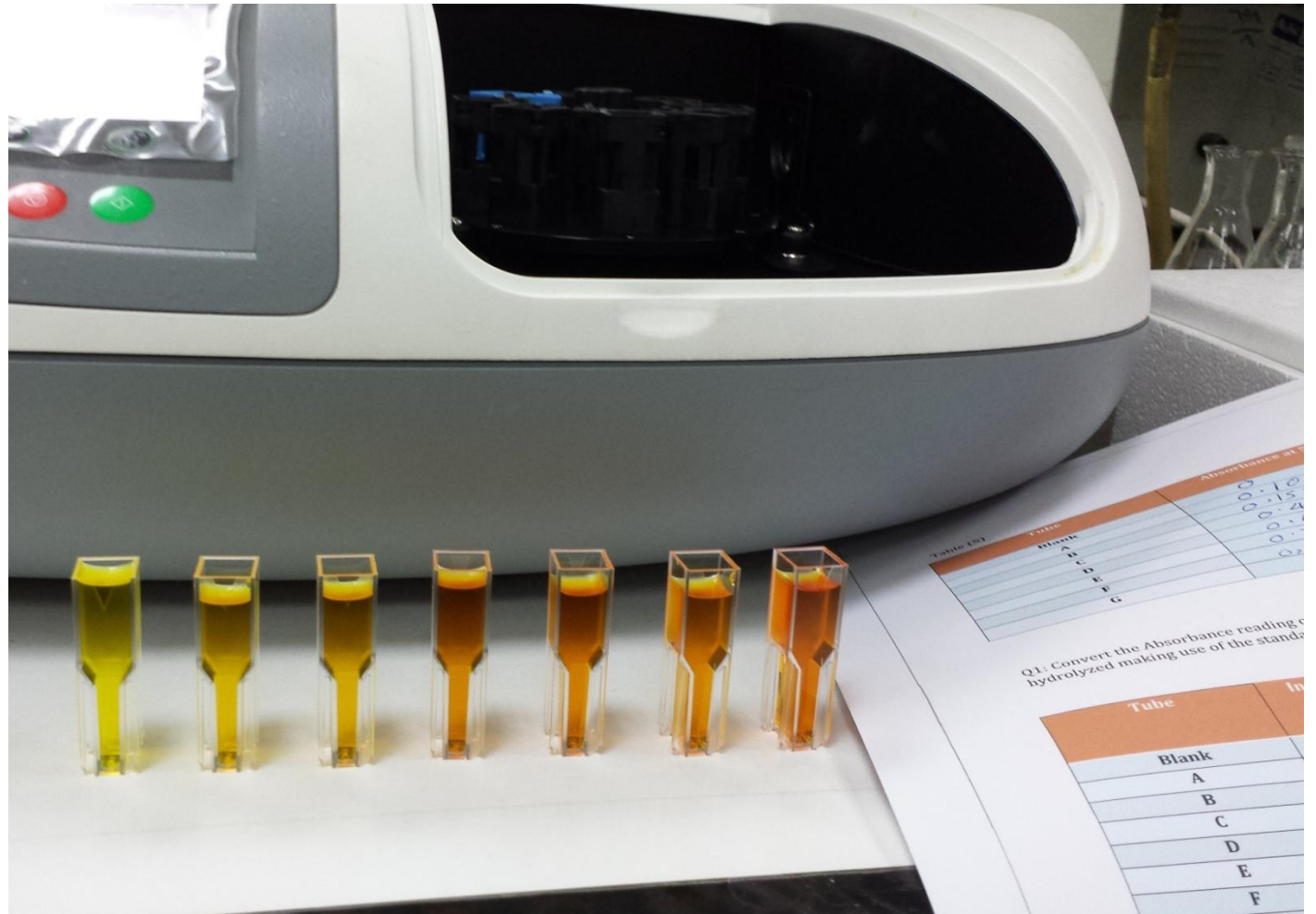
Material

Solutions :

- 0.05M Sodium Acetate buffer , pH 4.7 .
- 0.18 M Sucrose
- Reducing sugar (0.005M glucose + 0.005M fructose)
- β -Fructofuranosidase enzyme extract from yeast.
- DNS (dinitrosalicylic acid) Reagent .
- Sodium Bicarbonate .

Method :

As sheet provided



Result

- Part I:

Convert the Absorbance reading obtained to micromoles of sucrose hydrolyzed making use of the standard reducing sugars calibration curve , determine the initial velocity v_i for each tube. Table (1)

Tube	Absorbance 540nm	Concentration of sucrose *	μ moles of sucrose hydrolyzed
A			
B			
C			
D			
E			
F			

Note : the concentration of sucrose* is the accurate substrate concentration accounting for the diluting effect of both the acetate buffer and the enzyme , sucrose has been diluted by a factor of 0.655 in each tube .

Result:

- Part II:

Obtain the relationship between the initial velocity v_i and substrate concentration $[S]$, by drawing a graph between the initial velocity v_i and substrate concentration. **Determine the K_m value of the enzyme/ substrate pair . (Michaelis – Menten)** The results should provide classic Michaelis - Menten data from which approximations of V_{max} and K_m can be determined

Tube	Substrate Conc.	Velocity of reaction $\mu\text{moles of sucrose hydrolyzed/min}$
A		
B		
C		
D		
E		
F		

Results:

- Part III:

Obtain the relationship between the 1/initial velocity($1/v_i$) and 1/substrate concentration($1/ [S]$) .By taking the **double-reciprocal of the data in part II to find more exact values for K_m and V_{max} .**

Table (2)

Tube	Substrate Conc.	$1/(\text{Substrate Conc.})$ (x-values)	Velocity of reaction	$1/(\text{Velocity of reaction})$ (y- values)
A	
B	
C	
D	
E	
F	

Discussion:

- Comment on the results, patterns, shape of the curves and mention the reasons of shape obtained.
- Calculate the V_{max} and **Michaelis – Menten constant K_m** from the two graphs.
- Determine the K_m for **β -fructofuranosidase and sucrose** and comment on the affinity of sucrose to **β -fructofuranosidase**

Thank You