RESEARCH ARTICLE



Neuro-fuzzy model of homocysteine metabolism

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Abstract. In view of well-documented association of hyperhomocysteinaemia with a wide spectrum of diseases and higher incidence of vitamin deficiencies in Indians, we proposed a mathematical model to forecast the role of demographic and genetic variables in influencing homocysteine metabolism and investigated the influence of life style modulations in controlling homocysteine levels. Total plasma homocysteine levels were measured in fasting samples using reverse phase HPLC. Multiple linear regression (MLR) and neuro-fuzzy models were developed. The MLR model explained 64% variability in homocysteine, while the neuro-fuzzy model showed higher accuracy in predicting homocysteine with a mean absolute error of 0.00002 μ mol/L. Methylene tetrahydrofolate reductase (MTFR) C677T, 5-methyltetrahydrofolate homocysteine methyltransferase (MTR) A2756G and 5-methyltetrahydrofolate homocysteine methyltransferase reductase (MTRR) A66G were shown to be positively associatiated with homocysteine, while nonvegetarian diet, serine hydroxymethyltransferase 1 (SHMT1) C1420T and TYMS 5'-UTR 28 bp tandem repeat exhibited negative association with homocysteine. The protective role of SHMT1 C1420T was attributed to more H-bonding interactions in the mutant modelled compared to the wild type, as shown through *in silico* analysis. To conclude, polymorphisms in genes regulating remethylation of homocysteine strongly influence homocysteine levels. The restoration of one-carbon homeostasis by SHMT1 C1420T or increased flux of folate towards remethylation due to TYMS 5'-UTR 28 bp tandem repeat or nonvegetarian diet, and the towards remethylation due to TYMS 5'-UTR 28 bp tandem repeat or nonvegetarian diet can lower homocysteine levels.

Keywords. homocysteine; multiple linear regression; neuro-fuzzy design; diet.

Introduction

Homocysteine is a nondietary amino acid, which is the byproduct of methionine metabolism. Hyperhomocysteinaemia is documented as a potential risk factor for a wide spectrum of diseases such as recurrent pregnancy loss (Govindaiah *et al.* 2009), birth defects (Naushad *et al.* 2014a), psychiatric disorders (Moustafa *et al.* 2014), coronary artery diseases (Lakshmi *et al.* 2013), deep vein thrombosis (Naushad *et al.* 2007; Ghaznavi *et al.* 2015), Parkinson's disease (Kumudini *et al.* 2014; Kirbas *et al.* 2016), etc. Hyperhomocysteinaemia could occur due to genetic polymorphisms/mutations or cofactor deficiencies of the folate metabolic pathway.

The dietary source of folate is in the form of folyl polyglutamate and is converted to monoglutamates with the help of glutamate carboxy peptidase II (GCPII) in the intestine. With the help of folate reductase (FR), monoglutamates are reduced to dihydrofolate (DHF) and

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tetrahydrofolate (THF). Serine hydroxymethyl transferase 1 (SHMT1) present in the cytosol is a B_6 -dependent enzyme that catalyses the reversible conversion of serine and THF to glycine and 5,10-methylene THF; and irreversible conversion of 5,10-methylene THF to 5-formyl THF. Methylene tetrahydrofolate reductase (MTHFR) catalyses the flavin adenine dinucleotide (FAD)-dependent reduction of 5,10-methylene THF to 5-methyl THF. It also helps in the conversion of deoxy uridine monophosphate (dUMP) to deoxy thymidine monophosphate (dTMP) with the help of thymidylate synthase (TYMS). 5-Methyltetrahydrofolate-homocysteine methyltransferase (MTR) catalyses the conversion of homocysteine to methionine with 5-methyl THF as the substrate and methylcobalamin as the cofactor. 5-Methyltetrahydrofolate-homocysteine methyltransferase reductase (MTRR) catalyses the reductive methylation of cobalamin. Then, methionine is converted to S-adenosyl methionine (SAM), which donates methyl moiety to DNA, histones, catechol amines, etc. After the transfer of methyl groups, SAM is converted to Sadenosyl homocysteine (SAH), which on hydrolysis gives homocysteine.

The pathophysiology of hyperhomocysteinaemia was reported to be mediated through the following mechanisms: (i) auto-oxidation of homocysteine increasing free radical production (Zhang *et al.* 1998); (ii) the superoxide ion generated will remove nitric oxide from the circulation as peroxynitrite (Antoniades *et al.* 2006); (iii) homocysteine was shown to induce damage to endothelium (Pushpakumar *et al.* 2014); (iv) elevated homocysteine might be a surrogate marker of altered cellular methylation (Naushad *et al.* 2014); (v) it can promote hypercoagulable state by promoting procoagulants and inhibiting anticoagulants (Coppola *et al.* 2000) (figure 1).

Several putatively functional polymorphisms such as GCPII C1561T, RFC1 G80A, SHMT1 C1420T, TYMS 5'-UTR 28 bp tandem repeat, MTHFR C677T, MTR A2756G, MTRR A66G were reported in the folate metabolic pathway (Binia *et al.* 2014). These polymorphisms were shown to influence homocysteine levels and also contribute towards impaired DNA synthesis and DNA methylation. Deficiency of cofactors such as folic acid, B_2 , B_6 and B_{12} were reported to increase homocysteine (Sukla *et al.* 2013). Apart from these inherent variables, several other variables such as age, gender, hypertension, diabetes and consumption of tea, coffee, alcohol and smoking were also known to influence homocysteine (Landini *et al.* 2014).

In view of multi-system involvement of elevated homocysteine levels, we aimed to develop a mathematical model of homocysteine metabolism and utilize this model to understand the influence of environmental and genetic variables in modulating homocysteine levels. This will help in delineating whether lifestyle modulation can control homocysteine levels.



Figure 1. Homocysteine metabolic pathway. This scheme depicts the transfer of one-carbon moiety from one substrate to another in sequential reactions. Any perturbation in this pathway results in elevated homocysteine.

Method

Recruitment of subjects

Population-based controls in the age group of 20–75 yrs were recruited for this study. The inclusion criteria were: no history of any thyroid abnormality, no history of narcotic drug usage and no evidence of malignancy or inflammatory disease. We have recruited 85 subjects based on these criteria at Nizam's Institute of Medical Sciences, Hyderabad, India. The study protocol was approved by the Institutional Ethical committee of Nizam's Institute of Medical Sciences, Hyderabad, India. Informed consent was obtained from all the subjects.

Sample collection

Whole blood samples were collected in EDTA after overnight fasting. Plasma was separated immediately and stored at -80° C until further analysis. Buffycoat was used to extract DNA using phenol–chloroform extraction method following digestion with proteinase K.

Biochemical analysis

Plasma homocysteine and glutathione were investigated by reverse phase high performance liquid chromatography following precolumn derivatization with 4-fluro-7sulfobenzofurazan ammonium salt as described elsewhere (Ubbink *et al.* 1991).

Genetic analysis

GCPII C1561T, RFC1 G80A, SHMT1 C1420T, TYMS 5'-UTR 28 bp tandem repeat, MTHFR C677T, MTR A2756G and MTRR A66G were detected using polymerase chain reaction (PCR) restriction fragment length polymorphism (RFLP) and PCR-amplified fragment length polymorphism (AFLP) approaches as described earlier (Mohammad *et al.* 2011).

Multiple linear regression (MLR)

All genetic and environmental factors such as age, gender, GCPII C1561T, RFC1 G80A, SHMT1 C1420T, TYMS 5'-UTR 28 bp tandem repeat, MTHFR C677T, MTR A2756G and MTRR A66G were used as input variables and total plasma homocysteine was used as output variable to construct a MLR model.

Development of neuro-fuzzy logic model

For the development of neuro-fuzzy logic model, the input variables used were age, gender, diet, smoking, alcohol, diabetes mellitus, hypertension, GCPII C1561T,

dem repeat, MTHFR C677T, MTR A2756G and MTRR A66G, while total plasma homocysteine was used as the output variable. Fuzzy interference system (FIS) was generated using subclustering. The model was trained based on the data of 85 subjects. The FIS optimization for the training of the model was based on 'hybrid' method with error tolerance of 0.0001 and epochs of 3000. The training of the model was stopped when the mean absolute error was minimized. The performance of the model was ascertained by cross-validating the data as the testing and the checking data. The final model depicted the rules and surface plots representative of interactions between the

RFC1 G80A, SHMT1 C1420T, TYMS 5'-UTR 28 bp tan-

In silico analysis

variables.

The crystal structure of *Homo sapiens* recombinant serine hydroxymethyl transferase (PDB ID: 1BJ4) with a single chain of 480 residues was used as the template for this analysis. Using PyMOL software, the leucine residue at the 474th position was mutated to phenylalanine to generate the mutant protein structure. The binding affinity of three ligands, i.e., tetrahydrofolate (THF), methylene THF and formyl THF towards wild versus mutant was assessed using PyRx software. The chemical structures of ligands were obtained from drug bank. The atomic coordinates were generated using PyRx tool. The PDB coordinates of the protein and the ligands were optimized by Drug Discovery studio ver. 3.0. The binding energies were scored using Auto Dock vina option.

Results

25 20 15 15 10 5 6 CP II REC SHMT TYMS MTHER MS MTRR MALE The MLR equation explained 64% variability in homocysteine levels. Homocysteine levels were higher in men







Figure 3. Influence of diet on the association between genetic and demographic variables with homocysteine. This illustrates a comparative analysis between vegetarians and nonvegetarians (0, vegetarian diet; 1, nonvegetarian diet) demonstrating alterations in homocysteine with respect to age, MTR, MTHFR, RFC1 genotypes depicted as 0, 1 and 2 based on number of variant alleles. The plasma homocysteine levels were represented in μ moles/litre.

compared to women in all genotypes. The MTHFR C677T, MTR A2756G and MTRR A66G polymorphisms were shown to increase homocysteine (figure 2). The contribution of genetic and demographic factors towards homocysteine was depicted with the following equation:

Homocysteine =
$$(15.27) - (0.07^*\text{Age})$$

+ (4.93^*Gender)
- $(4.51^*\text{GCPII}) - (1.7^*\text{RFC})$
+ $(0.58^*\text{SHMT}) - (0.81^*\text{TYMS})$
+ (1.53^*MTHFR)
+ $(2.7^*\text{MTR}) + (2.19^*\text{MTRR}).$

The developed neuro-fuzzy logic model showed higher accuracy with mean absolute error of 0.000065. This model was developed to study gene–gene, gene–environment and gene–nutrient interactions. A positive association was observed between age and homocysteine level with respect to a vegetarian diet. Nonvegetarian diet showed a protective role (figure 3). An elevated level of homocysteine was observed in males compared to females (figure 4).



Figure 4. Association of age and gender with homocysteine. This illustrates changes in homocysteine in both genders across different age groups.

As shown in figure 5, MTHFR C677T was associated with increased homocysteine levels, while presence of



Figure 5. Counteracting interactions between MTHFR and TYMS in homocysteine metabolism. MTHFR C677T polymorphism was shown to increase homocysteine levels in the presence of TYMS 5'-UTR wild genotype. However, in the presence of 2R allele (1 and 2), MTHFR polymorphism will not exert homocysteine elevation.

TYMS 5'-UTR 28 bp tandem repeat negated this effect. The nonvegetarian diet was shown to decrease homocysteine levels induced by MTR A2756G (figure 3). MTHFR CT and TT genotypes were associated with elevated homocysteine in vegetarians. The nonvegetarian diet conferred protection against this polymorphism (figure 3). As shown in figure 3, RFC1 GA and AA genotypes increased homocysteine levels in vegetarians. The nonvegetarian diet conferred protection against elevated homocysteine. Since the SHMT1 C1420T has no appreciable impact on the homocysteine levels, an *in silico* analysis was performed to elucidate its functional impact. The binding affinities for the three ligands i.e., THF, MTHF and FTHF with wild versus mutant proteins were studied. Furthermore, H-bond interactions between the ligand and protein were elucidated.

The binding energies for wild and mutant proteins were -8.2 and -8.3 (kcal/mol), respectively, in the presence of THF as a ligand. When FTHF and MTHF were used as a ligand, these energies were similar i.e., -7.9 and -7.3 (kcal/mol), respectively. The interactions between the protein and ligand were viewed using PyMOL software. Though, the binding affinities of both wild and mutant types were the same, they differed in H-bonding interactions. A greater number of H-bond interactions were observed in the mutant than in the wild type (figure 6) protein in THF, while a greater number of H-bond interactions were observed in the wild type compared to the mutant in MTHF (figure 7).

Discussion

Hyperhomocysteinaemia is a well-documented risk factor for a variety of diseases affecting all age groups. Several



Figure 6. Interactions of THF with wild versus mutant proteins. The crystal structure of recombinant SHMT (PDB Id: 1BJ4) was used as a template and the leucine residue in 474th position was mutated to phenylalanine (L474F) using PyMOL. Docking was performed using PyRx, which showed similar binding affinities of wild and mutant proteins towards THF. However, H-bonding interactions were more in mutant protein than wild.



Figure 7. Interactions of MTHF with wild versus mutant proteins. The crystal structure of recombinant SHMT (PDB Id: 1BJ4) was used as a template and the leucine residue at 474th position was mutated to phenylalanine (L474F) using PyMOL. Docking was performed using PyRx, which showed similar binding affinities of wild and mutant proteins towards MTHF. However, H-bonding interactions were more frequent in the wild protein than in the mutant.

researchers have previously demonstrated the association of hyperhomocysteinaemia with the molecular pathophysiology of these diseases. However, due to complex gene-gene and gene-environment interactions, few studies were conducted to explain the elevation of homocysteine in terms of multilocus models. Mathematical models of homocysteine metabolism were proposed based on known reaction kinetics (Reed et al. 2004). An inverse association was reported between folate and homocysteine at very low concentrations of folate (Reed et al. 2006). The application of MLR approach to address the contribution of independent variables on a dependent variable is well accepted and this approach explained 64% variability in homocysteine. However, being a linear model, it is unlikely to take into account, complex interactions between the independent variables. We have overcome this limitation by applying neuro-fuzzy design, which is specifically capable of exploring bivariate interactions. The observations of this model showing positive association of MTHFR C677T and inverse association of TYMS 5'-UTR 28 bp tandem repeat and SHMT1 C1420T corroborated the mathematical model of folate metabolism (Ulrich et al. 2008), which showed increased homocysteine levels in the presence of decreased specific activities of MTHFR and SHMT; and increased specific activity of TYMS. Presence of promoter polymorphism in TYMS increases the flux of folate towards remethylation of homocysteine. Hence, even in the presence of MTHFR C677T, homocysteine will not be elevated. A similar mechanism was shown to confer protection against coronary artery disease (Vijaya Lakshmi et al. 2011).

A majority of Indians have been reported to have B_{12} deficiency by virtue of a vegetarian diet and hence, their homocysteine levels have been found to be higher than their counterparts across the globe (Refsum et al. 2001). To substantiate this observation, we compared changes in homocysteine levels with respect to different genotypes in vegetarians and nonvegetarians. We observed homocysteine lowering in nonvegetarians compared to vegetarians. The uptake of cobalamin and its reactivation were facilitated by specific protein-protein interactions between the MTRR FMN domain and MTR (Wolthers and Scrutton 2009). MTRR was proposed to serve as a molecular chaperone for MTR and also access aqua cobalamin reductase (Yamada et al. 2006). This hypothesis is substantiated by a study on Thai population, which showed protective role of nonvegetarian diet against MTHFR C677T induced hyperhomocysteinaemia (Kajanachumpol et al. 2013).

The association of MTHFR C677T with hyperhomocysteinaemia was consistent with the existing literature. Recently, we reported that because of this association, the frequency of this polymorphism is lower in South Indians due to adaptive developmental plasticity (Naushad *et al.* 2014b).

The association of RFC1 G80A with homocysteine corroborated our earlier observation showing an inverse association of this polymorphism with folate (Naushad *et al.* 2011). SHMT1 C1420T polymorphism showed no statistically significant association with the plasma homocysteine. To support this observation, *in silico* analysis was performed which suggested similar binding energy between the wild and mutant proteins for THF, FTHF

and methylene THF. However, the H-bond interactions were more in the mutant protein compared to the wild type for THF, while H-bond interactions were less in the mutant protein for MTHF compared to the wild type suggesting greater turnover of MTHF. This observation corroborated the studies on human and rabbit cytosolic SHMT, which showed no influence of mutation on the stability of SHMT or the rate of conversion from 5,10-methenyl THF to FTHF (Fu *et al.* 2005).

The major strength of the current study was the integration of MLR and neuro-fuzzy models to evaluate independent as well as bivariate interactions influencing homocysteine. Further, narrow mean absolute error increases the prediction accuracy of the model. The limitations of the current study were: (i) sample size; (ii) other lifestyle factors such as smoking, alcohol intake, consumption of tea and coffee, physical activity, etc., were not incorporated due to lack of quantitative information; and (iii) the estimations of folate and B_{12} were not performed due to technical constraints. Future studies are warranted on large ethnic groups and populations to increase the precision of these models.

To conclude, the MLR model of homocysteine explained 64% variability. The neuro-fuzzy model with narrow mean absolute error explored the interactions of remethylating polymorphisms with diet and demonstrated the protective role of nonvegetarian diet through lowering of homocysteine. Increased flux of folate either due to TYMS 5'-UTR 28 bp polymorphism or one-carbon homeostasis due to SHMT1 C1420T was shown to negate the homocysteine elevation mediated by MTHFR, MTR and MTRR polymorphisms.

References

- Antoniades C., Shirodaria C., Warrick N., Cai S., de Bono J., Lee J. et al. 2006 5-methyltetrahydrofolate rapidly improves endothelial function and decreases superoxide production in human vessels: effects on vascular tetrahydrobiopterin availability and endothelial nitric oxide synthase coupling. *Circulation* 114, 1193–1201.
- Binia A., Contreras A. V., Canizales-Quinteros S., Alonzo V. A., Tejero M. E. and Silva-Zolezzi I. 2014 Geographical and ethnic distribution of single nucleotide polymorphisms within genes of the folate/homocysteine pathway metabolism. *Genes Nutr.* 9, 421.
- Coppola A., Davi G., De Stefano V., Mancini F. P., Cerbone A. M. and Di Minno G. 2000 Homocysteine, coagulation, platelet function, and thrombosis. *Semin. Thromb. Hemost.* 26, 243–254.
- Fu T. F., Hunt S., Schirch V., Safo M. K. and Chen B. H. 2005 Properties of human and rabbit cytosolic serine hydroxymethyltransferase are changed by single nucleotide polymorphic mutations. *Arch. Biochem. Biophys.* **442**, 92–101.
- Ghaznavi H., Soheili Z., Samiei S. and Soltanpour M. S. 2015 Association of methylenetetrahydrofolate reductase C677T polymorphism with hyperhomocysteinemia and deep vein thrombosis in the Iranian population. *Vasc. Specialist Int.* **31**, 109–114.

- Govindaiah V., Naushad S. M., Prabhakara K., Krishna P. C. and Radha Rama Devi A. 2009 Association of parental hyperhomocysteinemia and C677T methylene tetrahydrofolate reductase (MTHFR) polymorphism with recurrent pregnancy loss. *Clin. Biochem.* 42, 380–386.
- Kajanachumpol S., Atamasirikul K. and Tantibhedhyangkul P. 2013 C677T methylene tetrahydrofolate reductase and plasma homocysteine levels among Thai vegans and omnivores. *Int. J. Vitam. Nutr. Res.* **83**, 86–91.
- Kirbas S., Kirbas A., Tufekci A., Cumhur Cure M., Cakmak S., Yazici T. *et al.* 2016 Serum levels of homocysteine, asymmetric dimethylarginine and nitric oxide in patients with Parkinson's disease. *Acta Clin. Belg.* **71**, 71–75.
- Kumudini N., Uma A., Naushad S. M., Mridula R., Borgohain R. and Kutala V. K. 2014 Association of seven functional polymorphisms of one-carbon metabolic pathway with total plasma homocysteine levels and susceptibility to Parkinson's disease among South Indians. *Neurosci. Lett.* 568, 1–5.
- Lakshmi S. V., Naushad S. M., Reddy C. A., Saumya K., Rao D. S., Kotamraju S. *et al.* 2013 Oxidative stress in coronary artery disease: epigenetic perspective. *Mol. Cell. Biochem.* 374, 203–211.
- Landini L. 2014 Modification of lifestyle factors are needed to improve the metabolic health of patients with cardiovascular disease risk. *Curr. Pharm. Des.* **20**, 6078–6088.
- Mohammad N. S., Yedluri R., Addepalli P., Gottumukkala S. R., Digumarti R. R. and Kutala V. K. 2011 Aberrations in onecarbon metabolism induce oxidative DNA damage in sporadic breast cancer. *Mol. Cell. Biochem.* 349, 159–167.
- Moustafa A. A., Hewedi D. H., Eissa A. M., Frydecka D. and Misiak B. 2014 Homocysteine levels in schizophrenia and affective disorders-focus on cognition. *Front. Behav. Neurosci.* 8, 343.
- Naushad S., Jamal N. J., Angalena R., Prasad C. K. and Devi A. R. 2007 Hyperhomocysteinemia and the compound heterozygous state for methylene tetrahydrofolate reductase are independent risk factors for deep vein thrombosis among South Indians. *Blood Coagul. Fibrinolysis* 18, 113–117.
- Naushad S. M., Pavani A., Digumarti R. R., Gottumukkala S. R. and Kutala V. K. 2011 Epistatic interactions between loci of one-carbon metabolism modulate susceptibility to breast cancer. *Mol. Biol. Rep.* 38, 4893–4901.
- Naushad S. M., Reddy C. A., Kumaraswami K., Divyya S., Kotamraju S., Gottumukkala S. R. *et al.* 2014a Impact of hype rhomocysteinemia on breast cancer initiation and progression: epigenetic perspective. *Cell Biochem. Biophys.* 68, 397–406.
- Naushad S. M., Krishnaprasad C. and Devi A. R. 2014b Adaptive developmental plasticity in methylene tetrahydrofolate reductase (MTHFR) C677T polymorphism limits its frequency in South Indians. *Mol. Biol. Rep.* 41, 3045–3050.
- Pushpakumar S., Kundu S. and Sen U. 2014 Endothelial dysfunction: the link between homocysteine and hydrogen sulfide. *Curr. Med. Chem.* 21, 3662–3672.
- Reed M. C., Nijhout H. F., Sparks R. and Ulrich C. M. 2004 A mathematical model of the methionine cycle. J. Theor. Biol. 226, 33–43.
- Reed M. C., Nijhout H. F., Neuhouser M. L., Gregory J. F. 3rd., Shane B., James S. J. *et al.* 2006 A mathematical model gives insights into nutritional and genetic aspects of folate-mediated one-carbon metabolism. *J. Nutr.* 136, 2653–2661.
- Refsum H., Yajnik C. S., Gadkari M., Schneede J., Vollset S. E., Orning L. *et al.* 2001 Hyperhomocysteinemia and eleva ted methylmalonic acid indicate a high prevalence of cobala min deficiency in Asian Indians. *Am. J. Clin. Nutr.* 74, 233– 241.
- Sukla K. K., Tiwari P. K., Kumar A. and Raman R. 2013 Low birthweight (LBW) and neonatal hyperbilirubinemia (NNH)

in an Indian cohort: association of homocysteine, its metabolic pathway genes and micronutrients as risk factors. *PLoS One* **8**, e71587.

- Ubbink J. B., Vermaak W. J. H. and Bissbort S. 1991 Rapid highperformance liquid chromatographic assay for total homocysteine levels in human serum. *J. Chromatogr.* **565**, 441– 446.
- Ulrich C. M., Neuhouser M., Liu A. Y., Boynton A., Gregory J. F. 3rd., Shane B. *et al.* 2008 Mathematical modeling of folate metabolism: predicted effects of genetic polymorphisms on mechanisms and biomarkers relevant to carcinogenesis. *Cancer Epidemiol. Biomarkers Prev.* **17**, 1822–1831.
- Vijaya Lakshmi S. V., Naushad S. M., Rupasree Y., Rao D. S. and Kutala V. K. 2011 Interactions of 5'-UTR thymidylate synthase polymorphism with $677C \rightarrow T$ methylene tetrahydro-

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folate reductase and $66A \rightarrow G$ methyltetrahydrofolate homocysteine methyl-transferase reductase polymorphisms determine susceptibility to coronary artery disease. *J. Atheroscler. Thromb.* **18**, 56–64.

- Wolthers K. R. and Scrutton N. S. 2009 Cobalamin uptake and reactivation occurs through specific protein interactions in the methionine synthase-methionine synthase reductase complex. *FEBS J.* 276, 1942–1951.
 Yamada K., Gravel R. A., Toraya T. and Matthews R. G. 2006
- Yamada K., Gravel R. A., Toraya T. and Matthews R. G. 2006 Human methionine synthase reductase is a molecular chaperone for human methionine synthase. *Proc. Natl. Acad. Sci.* USA 103, 9476–9481.
- Zhang F., Slungaard A., Vercellotti G. M. and Iadecola C. 1998 Superoxide-dependent cerebrovascular effects of homocysteine. Am. J. Physiol. 274, 1704–1711.